

Review Article

2018 | Volume 2 | Issue 2 | 39-60

Article Info

Open Access

Citation: Jarwar, A.H., Wang, S., Jarwar, Z.H., Ma, Q., Shuli, F., 2018. Use of Molecular Markers in Improvement of Cotton for Agronomic Traits. Int. J. Nanotechnol. Allied Sci., 2(2): 39-60.

Received: October 22, 2018

Accepted: November 10, 2018

Online first: November 23, 2018

Published: November 27, 2018

*Corresponding author: Fan Shuli; Email: Fsl427@126.com

Copyright: © 2018 PSM. This is an open access article distributed under the terms of the Creative Commons Attribution-Non Commercial 4.0 International License.



Scan QR code to see this publication on your mobile device.

Use of Molecular Markers in Improvement of Cotton for Agronomic Traits

Ameer Hussain Jarwar^{1, 3}, Xiaoyan Wang², Zaheer Hussain Jarwar⁴, Qifeng Ma¹*, Fan Shuli¹*

¹State Key Laboratory of Cotton Biology, Institute of Cotton Research of CAAS, Anyang 455000, China.

²Anyang Institute of Technology, College of Biology and Food Engineering, Anyang, 455000, China.

³Oil Seeds Section, Agriculture Research Institute, Tandojam, Sindh, Pakistan.

⁴Department of Commerce and Bussiness, Sindh University Jamshoro Sindh, Pakistan.

Abstract

Upland Cotton (Gossypium spp) is known as the largest natural fiber and vegetable oil source worldwide, to increase the superiority of agronomic traits withstand contrary abiotic and biotic stress in the field and fiber/yield qualities to fulfill all the necessity of advance spinning technology. Upland cotton increase through conventional plant breeding is a time consuming; present circumstance molecular markers established that effective tools to speed up the plant breeding programme for cotton improvement. Especially accentuate is given to application, obstacles, and perspectives of marker-assisted breeding since it appears to be more hopeful in falsify novel gene that are used in the cotton germplasm. The development of system quantitative breeding/genetics in molecular marker-helps for breeding programme would be necessary requirement to understand its role in cotton. While the same time, the function of genetic engineering and in vitromutagenesis cannot be used out in genetic melioration of cotton. In the demonstrate variety of molecular markers are useable, option of molecular marker depends on the users. The critique article gives a over view of versatile molecular markers used in cotton possess, Inter simple sequence repeats (ISSR), Random Amplified polymorphic DNA (RAPD), Amplified fragment length polymorphism (AFLP), Restriction fragment length polymorphism (RFLP), Sequence Related Amplified Polymorphism (SRAP), Single Nucleotide Polymorphism (SNP), and Simple sequence repeats (SSR), Above all molecular markers act a vital role in for betterment of crop improvement programme such as (a) Construction of linkage Map (b) Investigation of genetic diversity in cotton, (c) Marker Assisted Selection (MAS) (d) QTL investigation for agronomic and fiber related traits in cotton. Keywords: Molecular marker, Single nucleotide polymorphism, Cotton Improvement, GWAS, QTL.

PSM

International Journal of Nanotechnology and Allied Sciences

2018; 2(2): 39-60

INTRODUCTION

Upland cotton is a most crucial natural fiber crop cultivated in subtropical and moderate zones of atleast about 80 different countries of the world. The cotton fiber is used directly for raw material in textile, industrial and for cotton seed oil as a spin-off, (Li et al., 2014). Upland cotton incorporate atleast 50 approved species belongs to 8 genome groups (Wendal and Grover, 2015). From them 4 cotton species, such as G.hirsutum L, G.barbadence, G.herbaceum. and G.arborium had been cultivated for domesticated purpose. Whereas Gossypium hirsutum L. (2n=4x=52), the size of genome is 2.5 Gega bite; (li et al., 2015; 2014; Wendal and Grover, 2015). In addition, to specify the genetic base makes the productivity of cotton susceptible to attack different insect pests, any other fungal or any other disease. That is autochthonic to tropical and subtropical regions and existence cultivated on every continent of all over the world. In the world cotton crop is grown in an area of 35.1 million hectares producing 117 million bales with a productivity of 766 kg/ha (Dhruv, 2015). On the discipline that the quantitative traits divides into mortal genetic factors by finding deoxyribonucleic acid molecular marker closely connected with each others, that is very simple to control them expeditiously and this also helps to achieve the suitable results speedily and more exactly (Preetha and Rveendren 2008). Investigation of microsatellite or simple sequence repeats loci is high pragmatic due to of their duplicability transferability, codominant in nature (Ghaffari and Hasnaouri, 2013). In prescribe to compound all the favorable traits from different varieties are associate with wild species for evaluation of superior varieties through conventional breeding methods involve repeated backcrossing, testing and selfing they are time consuming and low exact processes as comparison to direct excerpt of plants basis on molecular procedures, (Preetha and Rveendren, 2008). SSR markers had been extremely used for the observation of genetic diversity, supervise of the introgression of novel alleles, quantity trait loci mapping, cultivars shelter, and choice of breeding (Rahman et al., 2008, Blenda et al., 2006, and Zhao et al., 2014.).

The immense majority of agronomical factors in upland cotton, as qualitative and quantitative traits, are controlled by many genes with little effects. Quantitative trait loci are effective tools which had been commonly used to detect the genetic architecture in quantitative factors. And it is also highly affected for using in marker assisted selection in crop breeding programmes. Diachronic markers, such as Simple sequence repeats (SSR), Amplified fragment length polymorphism (AFLP), and Restriction fragment length polymorphism (RFLP), had played a vital role in previous studies in QTL mapping in cotton. large number of interspecific and intraspecific genetic maps had been established and used in quantitative trait loci mapping by

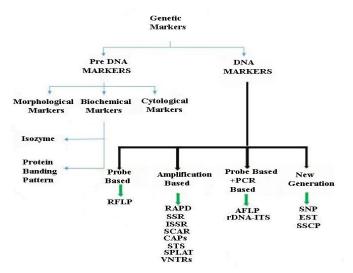
genetic linkage analysis in G.hirsutum L (Zhang et al., 2013; Yu et al., 2015; Wang et al., 2015a; Jamshed et al., 2016; and Fang et al., 2014). Linkage analysis can skim a minute quantitative trait loci and palace the quantitative trait loci in high regions (Powell and Mackay 2007; and Cavanagh et al., 2008). The using of high technology, such as single nucleotide polymorphism arrays and sequence, amend the firmness of molecular genetics maps and the accuracy of Quantitative trait loci mapping. high concentration of genetic linkage maps holding a high numbers of Single nucleotide polymorphisms or excision or addition polymorphism molecular markers had been observed on the basis of mixed type molecular markers. single nucleotide polymorphism are important locus (Wang et al., 2015; Yu et al., 2012; Zhang et al., 2016; Hulse-Kemp et al., 2015). Linkage with the physical genetic maps of G. hirsutum L. (Zhang et al., 2015; Li et al., 2015; Wang et al., 2012; Li et al., 2014). The tools which provides the content for fine mapping, identification of gene functional exploration, candidate gene, and Marker assisted selection (Zhu et al., 2016; Ma et al., 2016).

The GWAS is substitute tool for deleting quantitative trait loci and promising a genetic method to detection of some especial sites in plants parts (Saidou et al., 2014; Remington et al., 2001). Comparison to biparental genetic linkage mapping, Genome-wide association study had the reward of high-resolution, high price and no need for making a genetic population. Genome-wide association studies had been broadly used in studies of different plants, such as in the Rape and mustard, rice, maize. based on high database of discrepancy phenotypic and high concentration cultivars protecting the overall genome, the Genome-wide association studies is the effective and dominant for analyze genetic changes. Study the hindrance of population ascribes and surroundings changes, such as mixed linear model, general linear model, and Anderson test had been established using infrastructure, Relationship or other traits such as resultants changes to decrease mistakes (Yang et al., 2014; Liu et al., 2016). In last previous years, scientists have recognized many molecular markers loci colligate with plant characters, yield and fiber quality, resistance and yield by affiliation mapping using simple sequence repeats molecular markers (Liu et al., 2015; Nie et al., 2016; Abdurakhmonov et al., 2009), whileas, the molecular marker concentration and the representation population In this surveys was limited.

Tetraploid genome of upland cotton which is comparatively large and incorporate atleast 2200-3000 Mega byte of Deoxyribonucleic acid and morphological characterstics and RAPD, Crop Science, vol. no 36, no.1,pp. 186-192,1996). Interspecific Deoxyribonucleic acid polymorphism is too small in this species morphological characterstics and RAPD, Crop Science, vol, no,1, 36, pp. 186-192,1996, RFLP genetic diverseness in upland cotton,pp.81-101,Science Publishers, Enfield,

NH,USA,2001), that create it a ambitious crop for evaluation of markers. There is a huge demand for an high polymorphic marker on the condition of progress in crop creating using breeding is to be MAS plant breeding/techniques. In much quantity extraordinary reassessment had been written about the unlike classes of markers using in plants in construction of linkage map. The QTL analysis and MAS deoxyribonucleic acid markers, in Deoxyribonucleic acid-based molecular markers in plants species, pp. 39-57, Kluwer Academic Publishers, Dordrecht, the Netherland, 1994- marker engineering in Biotechnology and Molecular G.hirsutum, Biology reassessment, vol,3,no.2,pp. 32-45, 2008). The accusative of this review are as (a) analysis of the development of molecular marker technologies in cotton breeding and genetics, (b) genetic diverseness in the wild and cultivated cotton gene pools, and (c) An overview of QTL mapping and MAS activities in cotton.

Zhang et al., (2003) discovered eight markers connected with FS quantitative trait loci (QTLFS) which explicate more than 30% of the physical changes in a G.anomalum introgression line 7245. The quantitative trait loci was remains same in relative mapping of RIL and F2 population (Shen et al., 2005,2006,2007) it was offently used in Marker assisted selection plant breeding to increase fiber length (Guo et al., 2003). Conception of genetic molecular markers Mendal used phenotypic based genetic molecular markers research in his experiment estimate in the 19th century. Subsequently, genetic molecular markers formation of the hypothesis of genetic linkage map. (Reproduction plant cell, 2008 April; 27(4):617-31. The marker is delimited as a specific section of Deoxyribonucleic acid which is instance of the dissimilar at the genomic positions. The markers are not correlative with phonotypical manifestation of a trait. A schematic model of molecular markers in given in the figure 1.





Overview of marker methods

The release of Botstein *et al.*, 1990, genetic linkage maps using RFLP was the only first described marker method in the spying of Deoxsiribonucleic acid polymorphism.

The basic marker methods can be divided into two classes: (a) Polymerize chain reaction- based method (b) Non Polymerize chain reaction based method or interbreeding based methods and the other one is PCR-based method.

Polymerize chain reaction-based method

Later the conception of Polymerize chain reaction technology (Faloona & Mullis 1987), a wide No of ideas for propagation of markers found on polymerize chain reaction were elaborate, principally because of its evident easiness and high certainly of success. Using of unsequenced primers overcome the restriction of anterior sequence information for Polymerize chain reaction investigation and easy to the evolution of genetic molecular markers for a motley of purpose. Polymerize chain reaction-based methods can foster sub grouped into two sub groups: (1) haphazardly primed Polymerize chain reaction-based methods or sequence nonspecific methods and the other one is (2) Targeted sequence polymerize chain reactionbased methods.

DNA molecular markers techniques used in cotton Random Amplified Polymorphic DNA (RAPD)

Random amplified The polymorphism deoxsiribonucleic acid technique is different polymerize chain reaction amplification of genomic DNA. It deducts DNA polymorphosis create by rearrangement of excision at or among oligonucleotide primer connecting with genome using little random oligonucleotide order (Williams et al., 1991). Random Amplified Polymorphism DNA this is the basic Polymerize chain reaction-based molecular marker method it regards 10 bp primers (Williams et al., 1990). In this method has many reword over restriction fragment length polymorphism method such as no radioactive sleuthing, this also did not demand anterior sequence information, it mandatory very small measure of genomic DNA, observation chasteness and there is no need for costly tool beyond a thermo cycler and a transilluminator (Rafalski, 1997). The major disfavor of random amplified polymorphism DNA is a method had been used for high purpose in upland cotton possess appraisal of, genome mapping, diversity, and phylogenic studies(He et al., 2008, Rehman et al., 2002: Zhang et al., 2002: Bhat and Rana et al., 2004: Rahman et al., 2008), hereditary changes or diverseness studies (Chalmers et al., 1992, Xu et al., 2001; Tatineni et al., 1996; and Choudary et al., 2010), Deoxyribonucleic acid Multani et al., (1995) determines the association between the genotypes of various and same species Wajahatullah et al., (1997), he also estimate the

genetic/hereditary association between cotton varieties Shu et al., (2001), to recognized the guantitative trait loci for stomatal conductance Ulloaand Meredith, (2000), to manufacture linkage mapping and QTL analysis in G.hirsutum L (Wang et al., 2006, Lin et al., 2009, and Zhang et al., 2003). Random amplified polymorphic profile varies within and between laboratories because that is influenced by many factors such as DNA denseness, reproducibility of thermo cycler profiles, primer quality and density, selection of DNA polymerase, (Rafalski, 1997). Random amplified polymorphic was used to differentiate the cotton genotypes immune to aphid, mites, and jassids (Geng et al., 1995). Random amplified polymorphism DNA molecular marker R-6592 for male infertility gene had been established in G.hirsutum L (Lan et al., 1999). The DAF method regards using of individual arbitrary primers smaller than 10 nucleic acids for elaboration (Bassam and Caetano-Anolles, 1993).

Restriction fragment length polymorphism (RFLP)

Restriction fragment length polymorphism was the first kind of Deoxyribonucleic acid marker. In restriction fragment length polymorphism is discovered by hybridization a chemically labeled Deoxyribonucleic acid analysis. In southern blot compilation by restriction ecdonucleases, consequence in different fragment profile. That inhabitant to hybridization based molecular markers that are used for cloned DNA sequences to analysis a specific regions of the genome for changing that are realize as changes in the length of Deoxyribonucleic acid fragment produce by digestion with restriction ecdonucleases (Landry *et al.*, 1987).

RFLP was the initiative DNA molecular marker previously employed for crop betterment. Meredith et al., (1992) in a study of heterosis and varietal origins described that first restriction fragment length polymorphism rating in cotton (G.hirsutum L. Reinisch et al., 1994), described first RFLP based linkage map had 41 linkage with 4675 cm length with groups by using 700 RFLPs in cotton, (Chee et al., 2005, Paterson et al., 2003, Saranga et al., 2001: and Rong et al., 2004). The method is not broadly used because it is wasting of time, demand costly and radioactive/toxic reagents and expect high quantity and large quality genomic DNA. Yu et al., 1997, Used RFLP markers for genetic diversity studies in different G.hirsutum species. Reinisch et al., (1994) described that 46.2% of nuclear DNA analysis discover RFLPs among G.hirsutum and G. barbadense, 64% are highly dominant in nature than that many researchers were used that markers in constructing of linkage map in upland cotton. Wright et al., 1998, described utility of restriction fragment length polymorphism markers in marker help for selection (MAS) and RFLP connect to resistance allele for disease of bacterial blight was formalize. The demands of anterior sequence knowledge for probe multiplication improve the

complex methodology. These restrictions led to the formulation pair of small technically complex techniques called as polymerize chain reaction-based methods.

Single Nucleotide polymorphism (SNP) Markers and Population

Usually Genome-wide association studies had been used to diagrammatic quantitative factors in plants species (Lu *et al.*, 2015; Street and Ingvarsson, 2011; Zhao *et al.*, 2011; Crowell *et al.*, 2016; and Atwell *et al.*, 2010). The ability of Genome-wide association studies primarily consists of 4 components; the profuseness of genetic diverseness, acquired the maker concentration and stastical techniques. The *G.hirsutum* varieties accumulation originally from china had high levels of phenotypic and genotypic diversity. The accessions of 503 shroud the 5 cotton development zones in china. From them a little accessions were acquaint mainly from SN and from America, like as Stoneville and Coker Deltapine, that are the beginner of upland cotton breeding programmes in china and had implication contribute in cotton yield.

The closely high sample size guaranteed enough genetic variability, and the size was like to the Genomewide association studies used for A. thaliana (Zhao et al., 2005), Brassica napus (Xu et al., 2016), Oryza sativa (Famoso et al., 2011; Crowell et al., 2016), and Zea mays (Wen et al., 2014; Li et al., 2013a). The phonotypical variability associated with the environment upset the dependability of qualitative trait loci mapping. Multi environment plan and indifferent forecasting are practical manners to amend for this mistake. There are 4 sites for the factors analysis Hubei and Huanggang (HG, E115.77°, N29.45°); Yuanyang, Henan province (YY, E114.98°, N34.04°) Xijiang, Shihezi (SHZ, E84.92°, N42.28°); and korla, Xijiang (KX, E76.04°, N43.66°) are situated in the 3 major cotton zones in china. In improver, the 4 situations belong to 4 climatic divisions, BSK (cold and arid steppe), (hot temperate zones whereas the air is humid), Cwa (Desert & warm temperate regions), and Bwk (Arid desert and cold), (Du and Chen, 2006). Their are huge dissimilarities in geographic level and climate between these zones. There are sixteen agronomic traits having high phenotypic variability and stability herebility are suitable to expose their genetic foundation. The single nucleotide polymorphism array approach is dependable, effective and richly for genotyping. Upland cotton SNP 63K array, the first single nucleotide polymorphism for upland cotton, was established from pairs which symbolize inauspicious, possess G.hirsutum G.barbadence. G.tomentosum, G.mustelinum, G.armouriannum and G.long calyx (Hulse-Kemp et al., 2015). The array demonstrates high polymorphism which is (50.19%) in a cotton panel comparison with single sequence repeats markers (Wang et al., 2015, and Li et al., 2016). The common values concentration of polymorphic Single

nucleotide polymorphisms was ISNP/0.32 cM, equal to two hundred kilo byte in the phonotypical diagram that accomplishes the demand for Genome-wide association studies mapping.

Amplified fragment length polymorphism (AFLP)

The AFLP technique was previously discovered by a namely Vos et al., 1995; this method was based on three procedure regard three steps: (1) gel analysis amplified fragment, (2) Restriction of genome DNA and ligation of oligo nucleotide adopters, (3) pre and selective amplification of restriction fragment. This method compound dependability of restriction length fragment polymorphism with the simpleness of random amplified polymorphic DNA. Usually polymorphic fragment are perceived as presence or absence devising it a prevalent marker system, but in case of soya bean Maughan et al., 1996, detect co-dominant nature. This method can be automation and permit the Co-Occurrent analysis of many genetic loci per observations. Amplified Fragment Length Polymorphism (AFLP) creates more polymorphic loci per primer than SSRs RAPDs Maughan et al., (1996). AFLP is an effectual tool for the noticed the genetic diversity (Murtaza et al., 2006; Abdalla et al., 2002; Li et al., 2008; and Rana et al., 2005) finger printing studies, and tagging of agronomic, fiber and seed quality traits (Rakshit et al., 2010; Badigannavar et al., 2010; and Zhong et al., 2002). AFLP is most important method for mapping of gene studies due to their high copiousness and random distribution end to end the genome (Vos et al., 1995). An association relationship map of upland cotton was discovered using the AFLP and Random fragment Length DNA markers (Altaf et al., 1997). Amplified fragment length polymorphism molecular markers had also been used for constructing of linkage map and QTL analysis in addition with other markers Yu et al., 2007; Wang et al., 2006; Cuming et al., 2015; Lacape et al., 2009; and samer et al., 2015 and map saturation in G.hirsutum L. (Lacape et al., 2003, and Zhang et al., 2005).

Primers are also used for Amplified fragment length polymorphism analysis. Amplified fragment are perceived on denaturing polyacrylamide gels using an automation Amplified length fragment DNA sequence with the fragment option (Sun and Huang et al., 1999). The arrival of highly through orders technology, and high information on Deoxyribonucleic acid sequences for the genomes in different plant kingdoms had been created in Arabidopsis Genome opening 2000; Yu et al., 2002. Different crops species had been created and thousand of sequences had been comment as purport useable of genes using hefty bioinformatics tools. In prescribe to associate Deoxyribonucleic acid sequence knowledge with specific phenotypes.

The microsatellite based markers are small tandem reiterate or easy succession reiterate are flat repeating of too small 1-5 nucleotide theme, that happen as interspersed insistent components in many eukaryota genomes (Renz & Tautz 1984). Unevenness in the tandemly repetition units are primarily is due to maroon concurrently with Deoxyribonucleic acid reproduction where the reiterate permit matching via addition or deletion of iterates (Tautz and Schlotterer 1992). Equally slippage in reproduction is much similarly than point variations, microsatellite loci tend to be hyper variable.

The Polymerize chain reaction elaboration protocols used for microsatellites and is used sets or primers sets along one fluorolabelled or radiolabelled primer. Estimation of unmarkered Polymerize chain reaction product is carrying out using gel agarose. Using of microsatellite primers and laser sleuthing (such As., automation sequencer) in genotyping process had importantly amended (Wenz et al., 1998). Whileas, because of the maximum cost of the tag that is essential for to conduct by primers. Schuelke (2000) acquaint a process in which 3 primers are used for the elaboration rewords primer that demonstrates easy and little costly. Microsatellites are favorably famous genetic molecular markers due to of their predominant inheritance; high copiousness, extraordinary extent of allelic diverseness, and the simpleness of appraise simple sequence repeats size change by polymerize chain reaction along sets of primers. The duplicability of microsatellite markers that can be used expeditiously by many scientists in their laboratories to create coherent information (Maroof Saghai et al., 1994). The specific microsatellites-based molecular markers had been described from different plants species such as Lettuce (Lactuca sativa L.), and barley (Hordeum vulgare L.), (Saghai Maroof et al., 1994), (Van de wiel et al., 1999), and rice (Oryza sativa L.) (Tanksley and Wu et al., 1973).

Single nucleotide polymorphism (SNP)

Change of individual nucleotides (A, T, C, and G) in order of single genome is known as SNP (Agarwal et al., 2008). The Corn has single nucleotide polymorphism per 59-119 bp (Ching et al., 2002), whereas humans had an approximate 1 single nucleotide polymorphism 1,000 per bp (Sachidanandam et al., 2001). Single nucleotide polymorphisms discovery was high costly when based on sangar sequencing; so that have become costly effective with the use of different NGS technologies (Varshney et al., 2009, Metzker 2010). The Single nucleotide polymerize are commonly more predominant. In the coding or noncoding zones of the genome. Main advantage of single nucleotide polymorphism markers is to associate their simpleness of data management in addition with their flexibleness, speed and cost potency. It may present in coding, noncoding and intergenic zones of the genome, permit the espial of the genes is due to the changes in the order of nucleotides

Microsatellite Marker Methods

(Agarwal et al., 2008) and they are either non synonymous inside the coding zones. Synonymous variation can modify MRNA splicing that consequence the variation in the phenotype of a single (Richard et al., 1995). Bi-allelic single nucleotide polymorphism markers are straight forward to unite data across groups and make big databases of marker information, there are only two alleles per locus and different genotyping platforms will provide the like allele calls once suitable data had been accomplish. And the nucleotide polymorphism other single genotyping engineering like as genotyping by in order (Sonah et al., 2013; Elshire et al., 2011) can be simultaneously indentify and genotype the Single nucleotide polymorphism molecular markers or the identified markers can be used to sum more markers to genotyping assay (Davey et al., 2011).

Single nucleotide polymorphism markers are significant tool for linkage mapping, map based marker and cloning helps to selection due to the high level of polymorphism. Co-dominant nature of SNP creates those markers capable to differentiate the homozygous and heterozygous alleles (Shaheen *et al.*, 2009). Because of maximum polymorphism nature SNP was used to detect diversity, enactment, and mapping and for constructing of linkage map and QTL analysis in upland cotton (Hulse-kemp *et al.*, 2015, and Michael *et al.*, 2014).

In the recent, an international collaborative endeavor has developed 70K Single nucleotide polymorphism chip basis on genotyping essay unpublished data: http//www.cottongen.org/node/1287616. These huae throughput genotyping assays are the resources it can be used globally by private as well as public cotton plant breeders, Scienticists, and other geneticists to increase cotton genetic investigation. These are suitable to use their functions, including rapid recongnization of cultivars and constructing of ultra-high concentration genetic linkage map.

Progress in Marker Methods

In molecular markers the technical promotion as well genome based find had lead to the enhancement of molecular marker methods. The plant cell genomes such as mitochondrial DNA and chloroplast DNA had been progressively utilize to study genetic phylogenetic and structure association in plants. According to uniparental manner of contagion, mitochondrial genomes and chloplast showing different shape of genetic distinction comparison to nuclear genes (Provan et al., 1999). Therefore, overall apprehensions plant species of distinction and development, three interconnected genomes have been viewed, in improver to nuclear microsatellites, Molecular marker methods based on the mitochondrial and chloroplast had also been evolved. Improvement in orders engineering and accessibility of a growing number of expresses sequence tag sequence had create direct

investigation of genetic changes of the Deoxsiribonucleic acid orders (Sleimani *et al.*, 2003, Buetow *et al.*, 1999).

Actually the estimation of plastid organelle provide information of plants which are comprehensive to those receive from the nuclear genome. Various studies had described that chloroplast microsatellites lie on relatively small and number of mononucleotide extend such as (dA) n9 9dT) are omnipresent and polymorphic factors of chloroplast DNA (Powell et al., 1995). The genome based on markers visible genetic continuity and disparateness between taxa with rebuff physiological dissimilarities, which did not discover by nuclear deoxsiribonucleic acid markers as in the genetic breeding and interchange has vague, the manifest of previous demographic shape (Wolfe et al., 1987). Corresponding of orders in genome create it potential to comparison of allele in the plant species and analyze phylogenetic relationship in taxa which had deviate for hundreds years (Provan et al., 1999). The chloroplasts microsatellites are now decent steadfastly conventional as a huge-resolution tools for analyze shape of cytoplasmic change in a huge range of plants kingdom (Provan et al., 2001). The organelles are especially effectual markers for studying the pairing systems, gene stream for both seeds and pollen, and unapparent decent. The organelles microsatellite basis markers had been used for the espial of introgression and crossing (Bucci et al., 1998). The analysis of the genetic multifariousness (Clark et al., 2000) diverseness of plant species (Shaw et al., 2005; Parducci et al., 2001). The primer orders flanking organelles genomes. Generally endeavor to plan applicability primers to magnify chloroplast have consequence in a pair of microsatellite primers which purpose at amplifying cpsimple sequence reaction zones (Gardner and Weising 1999). Mainly the primer sets developed from T or A single nucleotide reiterate (n=10) indentified in the nicotiana organelles genome, were usable as genomic markers in the Cruciferae, Solanaceae and Actinidiaceae, (Staub and Chung 2003). The applicability primers for the elaboration of organelles in grasses family (Poaceae) had also been established (Provan et al., 2004).

A mitochondrial microsatellite in demarcation to carnal mtDNA calculates 319 MDa (Sedaroff et al., 1981). In improver huge size and shape, the to plant mtdeoxiribonuclei acid is qualified by molecular heterogeneousness discovered as sort of spherical chromosomes which change in shape and relative copiousness. In all plants species, genomes are commonly used for phyletic analysis because of a huge percentage of succession shakeup (Sederoff et al., 1981). Whatever, diversity mitochondrial associate to succession rearrangement testify helpful in population distinction of yearn and fir taxa (Sperisen et al., 2001; Soranzo et al., 1999). Mitochondrial reiterate had been used for sequestration population (Rajendrakumar et al., 2007).

Sequence Characterized Amplified Regions (SCAR)

In prescribe to use markers recognized by caprice marker analysis such as (RAPD, AFLP, etc.) for mapping based cloning, in individual locus moldiness be recognize unambiguously. Furthermore, the caprice marker methods are reasonable to vary in the reaction weather. In sequence to the space among the capability to receive associated markers to a gene of involvement in a short time and the usage of these markers for mapping cloning and for mundane screening processes, Sequence characterized Amplified regions marker method was discovered and employ. The SCAR are polymerize chain reaction based markers that symbolise genomic deoxyribonucleic acid fragment at genetically determine loci that are recognized by polymerize chain reaction elaboration using sequence particular oligonucleotide primers (McDermott et al., 1994; and paran and Michelmore 1993).

Etymologizing of Sequence characterized amplified regions regard propagate the amplified products of arbitrary marker methods that contrive particular primer sets of 15-30 bp that magnify individual main tie of the shapes alike propagate fragment. The polymorphism is either hold as the present or absent of elaboration of the tie or can involve in polymorphism change over prevalent caprice primed marker loci into predominant sequence characterized amplified regions markers. As sequence characterized Amplified regions are principally determined genetically, these are used for phonotypical as well as for genetic molecular markers. Predominant sequence characterized amplified regions are much informative are dominant primed markers; equally so these also be used to screen puddle genomic collections by polymerize chain reaction and for physiological map (Stephen and Chelkowski 2001), determining locus specificity (Michelmore and Paran 1993), equally relative map (Guo et al., 2003) and similar studies between comparative plant kingdoms.

Cleaved Amplified Polymorphic Sequence (CAPS)

Cleaved amplified polymorphic sequence marker technique which provide a technique to used the Deoxyribonucleic acid sequences of mapped Restriction Fragment Length polymorphism markers to evolve polymerize chain reaction based markers because obviate the tiresome DNA blotting (Konori Nitta and 2005). Consequently CAPS is also known as polymerizing chain reaction RFLP markers (Ausubel and Konieczny 1993). The cleaved amplified polymorphic sequences decode the RFLP caused by single base variation such as SNAs, insertion/deletions that alter restriction endo nuclease identification sites in PCR amplicons (Ausubel and Konieczny 1993; Chelkowski and Stephen 2001). Actually the CAPS assays the execute, by digest locus-specific polymerize chain reaction with single or many restriction enzymes, comply by detachment of the digested deoxsiribonucleic acid on gels polyacrylamide basis of

sequence information usable in databank of genomic Restriction amplified polymorphism DNA bands. Cleaved amplified polymorphic sequence analysis is versatile and can be connected with one SCP, RAPD, SSCP, SCAR, and ALFP, analysis to improve the possibleness of determination ,DNA polymorphisms, the cleaved amplified polymorphic sequence the molecular markers are predominant and locus particular and had been used to differentiate among plants which are heterozygous and homozygous for genes (Ausubel and Konieczny 1993). Therefore, Cleaved amplified polymorphic sequence demonstrate helpful as well as useful for genotyping, levels molecular identification studies (Spaniolas et al., 2006, Yu and Weiland 2003), the sequence based identification is not executable. The method is restricted by changes that make a restriction enzyme identification site. (Amasino 1998) suggest a discrepancy of the cleavage amplified polymorphic sequence method is known as dCAPS. The derived cleaved amplified polymorphism sequence analysis, a restriction enzyme acknowledgement site, that possess the single nucleotide polymorphism, is acquaint in the polymerize chain reaction product through primer consisting single or more than one not desirable to template deoxyribonucleic acid (Neff et al., 1998). That alter polymerize chain reaction product is than subjected to present or absent of the single nucleotide polymorphism is determined by the consequence restriction shape. This method is cheaper and easy, and used the omnipresent engineering of polymerize chain reaction, agarose gel and restriction digestion analysis. This method demonstrates helpful for variation or segregating population and based on propagating a copy of genotypes in plants species (Haliassos et al., 1989).

Randomly Amplified Microsatellite Polymorphisms (RAMP)

Randomly amplified microsatellite polymorphism (RAMP) microsatellite based markers demonstrate a high range of allelic polymorphism but these are too much costly and labour expenditures. Meanwhile in the other side Random amplified polymorphism DNA markers are cheaper but display small range of allelic polymorphism. To correct for the failing of both two sources, a method namely as randomly amplified microsatellite polymorphism was established (Wu et al., 1994). This method regard a radio labeled primer lie of a 3 repeats and 5 anchors which is used for to magnify genomic DNA in the present or absent of Random amplified polymorphism DNA primers. So the boiling temperatures of the anchored primers are offently 10-15C higher than the Random amplified polymorphism DNA primers therefore at high temper expeditiously, whilemean in polymerize chain reaction oscillation at small annealing temperatures both anchored microsatellite and Random amplified polymorphism DNA primers would temper. The polymerize chain reaction plan was alter like

as that there is switching among low and high tempering temperatures in the reaction. Many fragment receive with Randomly amplified microsatellite polymorphism primers alone vanish when Random amplified polymorphism DNA primers are enclosed, and different shapes are receive with the similar RAMP primers and different Random amplified polymorphism DNA showing that restriction amplified polymorphism DNA primers contend with Randomly Amplified Microsatellite Polymorphisms primer in the low tempering temperature cycle. Randomly amplified microsatellite polymorphisms had been used in studies of the cultivars of peach (Cheng *et al.*, 2001).

Target Region Amplification Polymorphism (TRAP)

TRAP method (Vick and Hu 2003) is an efficient and rapid polymerize chain reaction-based method that is used for bioinformatics tools and explicit sequence tag (EST) database knowledge to create polymorphic markers. These methods use two primers (nucleotide 18 in ranges) to create markers. Among of the primers, the rigid primer, is intentional from the aim explicit sequence tag sequence in the database; and the other one primer with At- or Gc to temper with a exon or intron. While the target region amplification polymorphism method used to create markers for specific gene sequence, that is helpful for germplasm genotyping and creating markers colligate with suitable for agronomic factors, plant breeding for marker assisted breeding (Hu et al., 2005). The technique had been efficaciously used in fingerprinting in cabbage genotypes (Hu et al., 2005), in calculating genetic diversity to mapping quantitative trait loci in wheat intervarietals recombinant inbred species (Liu et al., 2005).

Transposable elements-based molecular markers

Actually the transposable elements are the mobile genetic components able to modification their level in the genome. These are observed proximally previous 60 years in corn. There are two wider sections of transposable components. each with characterstics properties (Finnergan 1988). In section I or retro elements, such as retrotransposons, small interspaced nuclear components, and large interspaced nuclear components, these component encoded mRNA arbitrates markers. So in every transposition event generate a duplicate copy of the transposon whereas the original duplicate copy remains entire at the presenter site. In demarcation, section ii it lies of Deoxsiribonucleic acid transposon that changes their emplacement by a 'paste or cut' mechanics (Grzebelus 2006). That means they expunge themselves from the presenter site and regenerate themselves at the accepter site.

Based on the diagrammatical features, transposons are be foster sub classified into sub sections, super families, subfamilies, and families' basis on the structure and orientation of clear reading produced upon insertion (Grzebelus 2006). Large genomes and the retrotransposons are the basic unit of insistent deoxyribonucleic acid (Bennetzen and Kumar 1999) incorporates 35-55% from the total genome. Based on geomorphologic administration and fatty acid similarities encoded rearward between their transcriptases, retrotransposons also split into 3 classes. The long terminal repeats are mention to as the copia and gypsy like retrotransposons. And the 3rd one part of retrotransposons, line-1 like or non-long terminal the repeats retrotransposons, deficiency of TR and coded proteins with importantly few similarity to the retrovirus.

The retrotransposons Gypsy like (Suoniemi et al., 1998) and copia like (Kumar et al., 1996; Voytas et al., 1992) are demonstrated all over the kingdom plantae. Retrotransposons provide a very good chance to evolve marker system (Kalender et al., 1999) explained, protected orders and new inspectional polymorphisms created by replication ally are the predominant members. And the new intromission assist organizing intromission events temporally in a decent (Shimamura et al., 1997) so that are helpful for a ascertain pedigree and phylogenetic (Hafez et al., 2006). In the inter retrotransposons amplified polymorphism. DNA among retrotransposons microsatellite magnifies polymorphism (REMAP) regard elaboration of fragment that lies between retrotransposon intromission location and a microsatellite site. RBIP discover loci filled by or hollow of a retrotransposon.

RNA-based molecular markers

The SSCP analysis of RT-Polymerize chain reaction merchandize are be used to appraise the manifestation condition (bearing and comparative quantity) of much alike homologus gene sets from a polyploidy genome. Repeated trial demonstrates that cDNA-SSCP dependably split copy transcripts with 98% sequence observed (Adam and Cronn 2003). This method had been used to increase remarkable penetration into the globular frequency of still in natural and synthetic polyploidy.

Ribonucleic acids fingerprinting by randomly primed polymerize chain reaction the RAP-polymerized chain reaction methods (Welsh et al., 1992) require fingerprinting of ribonucleic acid populations using randomly select primer at small Strictness for 1st and 2nd strand cDNA synthesis comply by polymerize chain reaction elaboration of cDNA population. This method demand nano grams of overall ribonucleic acid and is insensible by small percentage of genomic Deoxyribonucleic acid pollution. Derivative polymerize chain reaction fingerprints are discovered for RNAs among the similar tissue isolated from different mortal and for RNAs among different tissues from the similar mortal. The single-specific different disclose because of to sequence polymorphisms and these are helpful for genetic map of genes. These tissue-particular different discover are helpful as well as for study of different

gene manifestation. CDNA-Amplified fragment length polymorphism is a novel ribonucleic acid fingerprinting method to exhibit different explicit genomes (Bachem et al., 1996). The way possess digestion of cDNAs by two restriction enzymes comply by ligation of oligo nucleotide adaptors and polymerize chain reaction elaboration used primers complementary to the adopter succession with increasing the select nucleotides (Bachem et al., 1998). The cDNA-Amplified fragment length polymorphism method is a high rigorous and reproducible than RAP-Polymerize chain reaction (Pardee and Liang 1992). In demarcation to interbreeding-based methods, like as cDNA micro arrays, cDNA-Amplified fragment length fragment can differentiate among high homogenous allele from single gene species. So no demand of any pre-existing orders knowledge in cDNA- Amplified fragment length polymorphism, so that is meaningful as a tool for the designation of novel process colligate alleles (Yaa et al., 2007; Van der Hoveven et al., 1996; and Akihiro et al., 2006). Designation of stress-regulated alleles had been a most important procedure of cDNA- Amplified fragment length polymorphism (Mao et al., 2004).

Impact of Molecular Marker Methods

When the coming of marker, it is present potential to create direct inferences about genetic diverseness and relationships between organisms at the Deoxyribonucleic acid level without the contradictory effects of the surroundings to pedigree records. The genetic analysis of living organism's population and species for taxonomic, ecological studies enormously profited from the evolution of different markers methods. An oblivious trouble that commonly originate is, how to select the superlative proper DNA marker between the infinite of different markers methods. Generally, the selection of a marker method had to be a compromise among dependability and simple investigation, stastical confidence and ability of discloser polymorphisms (Table 1).

The start molecular marker technology which was used for physiological map of plant genome was Restriction fragment length polymorphism. The method demand anterior sequence knowledge and is costly. With the innovation of Polymerize chain reaction engineering, marker methods like as AP-PCR, Amplified fragment length polymerization, Restriction amplified polymerization DNA, were established. These methods are so fast, cheaper and don't demand anterior sequence information. Methods like as AFLP and RAPD had been every day used for genetics population (Althoff et al., 2007) and for crop breeding progremmes. These can be used for marking a physiological factor to a genetic factor (Agarwal et al., 1992). In same manner to change haphazardly primed Polymerize chain reaction products in genomics, The sequence characterized amplified regions method was intentional. Microsatellite marker method used the inters as

well as intra mortal changes in simple sequence reiterate region for fingerprinting analysis.

Many marker methods are used to appraise the genetic diversity to construct a physical map of the genome being studies. Physical map of the connected markers assist in affiliation of the physical space to the genetical space among them. Association of the form to constructing of genetic linkage maps by emplacement of various polygenic and monogenic factors to particular zones of the plant species. The restricted genomes are kilo bases to several mega bases. After that the apart by pulse field gel electrophoresis and investigated by southern blot interbreeding using analysis incorporate of relatively linked markers. If analysis symbolize two markers hybridizes to the similar fragment, and the size of fragment is taken to be high space between the two markers.

A Biotic Stress

Cotton yield is damaged by different abiotic factors that cause about 72% decrease of cotton yield (Saranga et al., 2009). Between them salinity and drought these two factors are that effectually effect on cotton production, and it's a big problem so generate improved tolerance cotton verities against these stresses. However, some MAPK cascades factors had been described in different crops. Two Kinases, such as GhMKKI and GHMKKS had been described in accentuate resistance in upland cotton (Lu et al., 2013, Zhang et al., 2012). Over dominant of GhMKKI in nicotiana amend which toleratrate to drought and salt stresses, showing an intensified scavenging capacity and highly minded behavior of antioxidant enzymes (Lu et al., 2013). In other studies, a drought hypersensitive variant of a positive MAPK kinase gene had been recognize in Oryza sativa (Ning et al., 2010). Conversely, the oscpk 12 changes in RNAi plants were highly sensible to high salinity and accumulated vigorous water compare to wild species plants (Asano et al., 2012). The protected WRKY domain act significant rules in different morphological methods by connecting to W-box the booster zones of target alleles (Somssich and Ulker 2004; Wang et al., 2015; Ruston et al., 2010). Described a stress reactive GmWRKY27, WRKY gene, reduce ROS level and enhance drought and salt tolerance in transgenic Glycine max roots. The GmWRKY27 acts with GmMYB174, that is turn, acts in performing to decrease booster activity and gene manifestation of GmNAC29 (Wang et al., 2015). Subsequent analysis demonstrates that GhWRKY17 includes in stress responses in ABA (Sun et al., 2015, and Yan et al., 2014). Glycine max GmNAC2, NAC TF, was established as a negative regulator in the abiotic stress, (Ramegowda et al., 2012, and Jin et al., 2013). Isolated a stress reactive NAC gene, EcNACL, from finger Pennisetum glaucum. In upland cotton such as MAP3k gene extremely regulates defence but arbitrate decrease

tolerance to Abiotic and biotic stress in transgenic tabacum (Chen *et al.*, 2015).

Biotic factors

The biotic stresses are due to disease, weeds, pathogens, and insects hold with different levels of intensity, they generally decreased the plant population in crops (Borem and Fritsche-Neto, 2012). Between the various biotic factors, the cotton breeding against pest immune remains the basic aim. Due to shortage of

resistant cotton genotypes creates a most severe disease to regulate cotton yield (Chang *et al.*, 2008). And the cotton leaf curl virus is also one of them a serious disease in cotton that because decrease or reduces cotton production, it has been reported that resistant against cotton leaf curl virus is bestow one superior and two dominant gene by (Rahman *et al.*, 2005). To stimulate the resistance against insects, using of insect herbivores to resistance against insect herbivores.

	Abundance	Reproducibility	Degree of	Locus	Technical	Quantity DNA	Major
			polymorphism	specificity	requirement	required	application
RFLP	High	High	Medium	Yes	High	High	Physical
							mapping
PAPD	High	Low	Medium	No	Low	Low	Gene Tagging
SSR	Medium	Medium	Medium	No	Medium	Low	Genetic
							Diversity
SSCP	Low	Medium	Low	Yes	Medium	Low	SNP Mapping
CAPS	Low	High	Low	Yes	High	Low	Allelic Diversity
SCAR	Low	High	Medium	Yes	Medium	Low	Gene tagging &
							Phy- Map
AFLP	High	High	Medium	No	Medium	Medium	Gene tagging
IRAP/	High	High	Medium	Yes	High	Low	Genetic
PREMAP	-	_			-		Diversity
RAMPO	Medium	Medium	Medium	Yes	High	Low	Genetic
					-		Diversity

Table 1 comparing of diverse aspects of frequently used molecular marker methods.

Restriction fragment length polymorphism (RLFP), Random amplified polymorphic DNA (RAPD), Simple sequence repeats (SSR), Single strand conformational Polymorphism (SSCP), Cleaved amplified polymorphic sequence (CAPS), Sequence characterized amplified region (SCAR), Amplified fragment length polymorphism (AFLP), IRAP/REMAP inter-retro transposon amplified polymorphism Retrotransposon-microsatellite amplified polymorphism (IRAP/REMAP).

Major applications of molecular markers for betterment of cotton improvement

Genetic diversity surveys in upland cotton

Breeding program achiever depends on the apprehension of genetic diversity among and between genetic available germplasm and enable for breeders to select parental sources which will create diverse populations for selection. Portrayal of genetic similarity between genotypes is sources to select parental combining for keep genetic diversity in a plant breeding progamme (Beccelaere *et al.*, 2005). The information of genetic relationships between plant genotypes used to know the complexity, to detect the comparison in available genotypes and to make up utile conservation plants (Dahab *et al.*, 2013). Therefore, rating based on the markers that give meaningful that is useful in the evolution of new genotypes. There are many genetic diversity surveys had been carry out in cotton by using of different markers methods such as, RAPD (Choudary *et* *al.*, 2010; Xu *et al.*, 2001), AFLP (Rana *et al.*, 2004; Lie *et al.*, 2008; Abdalla *et al.*, 2001) simple sequence repeats (Arunita *et al.*, 2010; Qayyum *et al.*, 2009). A overview of some published genetic diversity surveys by using molecular markers is described in Table-1.

Genetic Linkage map building in upland cotton

Genetic linkage mapping (is cognize as meiotic mapping or linkage mapping) mentions to the finding of the relative level and space among markers with chromosomes. The genetic map space among two markers is described as the mean No of recombination happens; regard a given chromatid, in this zone per meiosis. Genetic linkage maps are basic for the localization of genes conferring biotic and abiotic stress tolerance. The genetic linkage map bases on markers have much reward over classical maps. Genetic mapping can be developed by different mapping populations, but popularly F2, backcross and recombinant inbred lines these three populations were used for

2018; 2(2): 39-60

constructing of genetic linkage map in plants (Paterson, 1996). The molecular map of the upland cotton genome was first manufactured by using 705 Restriction fragment length polymorphism loci and separate into 41 linkage groups (Reinish *et al.*, 1994). Many more cotton molecular maps had been constructed and published. A summary of published genetic linkage maps in upland cotton is given in (Table-2).

QTL Map for Fiber and Yield Quality Trait Contributing in Cotton

The zones in genomes to have genes connected with a qualitative trait are called as QTLs (Collard *et al.*, 2005), and the producer of evolving genetic linkage maps and playacting QTLs analysis is mentioned to as quantitative trait loci mapping (Paterson *et al.*, 1996). The quantitative trait loci analysis supports on the principal of identification a linkage between genotype and phenotype of markers. The QTLs identification in *G.hirsutum L* using different marker engineering is listed in table-3. They identified the QTLs are the new approach to boost up the cotton betterment through marker assisted option.

Marker assisted selection (MAS)

The Marker Assisted Selection is a technique through which a phenotype is selected on the behalf of genotype of a molecular marker (Collard et al., 2005). Once the markers associated to the genes had been discovered, cotton breeders use specific DNA molecular markers to identify the plants carried out the genes (Young et al., 1996). The potency and the cost of molecular marker assisted selection are determined by the marker method; thus, it must be selecting cautiously (Young et al., 1996). During the previous two decades, Restriction Amplified polymorphism DNA method had been used for Marker assisted selection for acquiring the glandless seeds and glanded plants. That was exposed that DNA markers associated to the major quantitative trait loci (QTLFSI) for the fiber strength could be used in marker assisted selection to growth fiber strength of commercial genotypes in segregating population (Zhang et al., 2003). SSR markers namely CIR 316 closely connects to root knot nematode (RKN) resistant region on chromosome 11 and BNL 3661 marker closely linked to RKN resistant region on chromosome 14. Jenkins et al., (2012) by used these SSR markers selected 11 homozygous plants for chromosome 11 and 14 from f2 population derived from RKN resistant genotype M 240 RNR x Susceptible cultivars FM 966 alternatively of waiting up to F6-F8 through conventional breeding methods. That selects the plant confirmed resistance against the RKN. Upland cotton it is necessary to recognized specific genes for specific traits such as strength fineness, fiber length. Etc., to compound those genes from different genotypes through marker assisted selection.

Table 2. An overview of	genetic diversit	y survey in u	pland cotton b	y using	g molecular markers.
-------------------------	------------------	---------------	----------------	---------	----------------------

S. No.	Country	Population type	Markers used	References
1	USA	24 lines of cotton	270 SNP loci and 92 Indel	Van <i>et al.,</i> 2009
2	India	24 lines of G. hirsutum L	6 AFLP primers	Rana <i>et al.,</i> 2005
3	Pakistan	31 Gossypium species 3 subspecies and 1 interspecific hybrid	45 RAPD primers	Khan <i>et al.,</i> 2000
4	USA	24 cultivars of G. hirsutum	88 SSR primers	Zhang <i>et al.,</i> 2005

Table 2a. An Overview of published genetic linkage maps in upland cotton G.hirsutum L.

S. No.	Cross parents	Mapping population		Markers	No. of mapped loci	Map Len gth (cM)	No.of LGs	Reference
	Interspecific crosses	Туре	Size					
1	Gh(palmeri) × Gb (K101)	F2	57	RFLP	705	4675	41	Reinisch <i>et al.,</i> 1994
2	Gh (CAMD-E) × Gb	F2	271	RFLP	261	3767	27	Jiang et al., 1998



2018; 2(2): 39-60

	(Sea Island Seaberry)							
3	Gh(Deltapine 61) × Gb (Sea IslandSeaberry)	F2	180	RFLP	-	3664	26	Jiang <i>et al.,</i> 2000
4	Gh(TM1) × Gb (3-79)	F2	171	RFLP,RAPDan dSSR	-	4766	50	Kohel <i>et al.,</i> 2001
5	Gh(Siv'on) × Gb (F- 177)	F2	430	RFLP	253	-	-	Saranga <i>et al.,</i> 2001
6	Gh(Siv'on) × Gb (F- 177)	F3	208	RFLP	-	-	-	Paterson et al., 2003
7	(Gh(TM 1) × Gb (Hai7124)) × TM1	BC1F1	140	EST-SSR	624	5644.3	34	Han <i>et al.,</i> 2004;2006
8	Gh (Acala 44) × Gb (Pima S7)	F2	94	AFLP,SSR,and RFLP	392	3287	42	Mei <i>et al.,</i> 2004
9	Gh(Palmeri) × Gb (K101)	F2	57	RFLP	2584	4447.9	26	Rong <i>et al.,</i> 2004
10	(Gh(Tamcot 2111) × Gb (Pima S6))×Tamcot 2111	BC3F2	3662	RFLP	-	-	-	Chee <i>et al.,</i> 2005
11	(Gh(Guazuncho 2) × Gb (VH8)) × Guazuncho 2	BC1 and BC2	200	SSR and RFLP	1306	5597	26	Lacape <i>et al.,</i> 2003; Lacape <i>et al.,</i> 2005
12	Gh(Handan208) × Gb (Pima90)	F2 and F2:3	69	SSR,SRAP,RA PD, and REMAPs	1029	5472.3	26	Lin et al., 2005; He et al., 2007
13	Gh(TM1) × Gb (Pima 3- 79)	RILs	183	EST-SSR	193	1277	19+11L G	Park <i>et al.,</i> 2005
14	Gh(7235) × Gb(TM-1)	F2 and F2:3	163	SSR	86	666.7	21	Shen <i>et al.,</i> 2005
15	Gh(TM1) × Gb (3-79)	RILs	183	SSR	433	2126.3	46	Frelichowski <i>et al.,</i> 2006
16	Gh(CRI36) × Gb (Hai7124)	F2	186	SSR, TRAP, SRAP, and AFLP	1097	4536.7	35	Yu <i>et al.,</i> 2007
17	Gh(Handan 208) × Gb (Pima 90)	RILs	121	SSR	-	5472.3	26	He <i>et al.,</i> 2008
18	Gh(Guazuncho 2) × Gb (VH8-4602)	RILs	140	SSR and AFLP	800	2044	26	Lacape <i>et al.,</i> 2009
19	(Gh(KC3) × Gb (Suvin)) × KC3	BC1F1	62	SSR	57	911.6	19	Santoshkumarr <i>et al.,</i> 2010
20	Gh(TM1) × Gb (3-79)	RILs	186	SSR and SNP	2072	3380	26	Yu et al., 2012
21	Gh(SG 747) × Gb (Giza 75)	BILs	146	SSR	392	2,895	26	Yu <i>et al.,</i> 2013
22	Gh(TM-1) × Gb (NM24016)	RILs	98	SSR and SNP	841	2061	26	Michael <i>et al.,</i> 2014
23	Gb.doubled haploid line 3-79 x G. hirsutumcv. Texas Marker- 1	F2	118	SNP	19,198	4,439.6	0.23	Hulse-Kemp <i>et al.,</i> 2015
24	Giza 45 (G. barbadense) xTamcot Luxor (G. hirsutum)	F2	60	AFLP,SSR, EST-SSR	210	3503.80	26	Samer <i>et al.,</i> 2015
25	Gh(TMS22) × G.tomentosum(WT936)	F2	82	SSR	589	4259.4	52	Westengen <i>et al.,</i> 2005



Table 2b. An overview of published genetic linkage maps in upland cotton.

	Interspecific crosses	Туре	Size	Markers	No. of mapped loci	Map Length (cM)	No.of LGs	Reference
1	Gh(HS46) × Gh(MARCABUCAG8U S-1-88) F2 and F3 96 RFLP 120 865 31	F2 and F3	96	RFLP	120	865	31	Shappley <i>et al.,</i> 1998
2	Gh× Gh	F2:3	569	RFLP	284	1502.6	47	Ulloa <i>et al.,</i> 2002
3	Gh(TM 1) × G.anomalum(7235)	F2 and F3	186	SSR and RAPD	-	-	-	Zhang <i>et al.,</i> 2003
4	Gh (Handan208) × Gh(Pima90)	F2	129	SRAP	237	3030.7	39	Lin <i>et al.,</i> 2005
5	Gh (Acala 44) × Gb(Pima S7)	F2	94	AFLP, SSR, and RFLP	392	3287	42	Mei <i>et al.,</i> 2004
6	G.trilobum(Skovsted)× G. raimondii(Ulbr)	F2	62	RFLP	763	1493.3	13	Rong <i>et al.,</i> 2004
7	Gh(Yumian 1) × Gh(T586)	F2 and F2:3	117	SSR and AFLP	70	525	20	Zhang <i>et al.,</i> 2005
8	Gh(TM1) × Gh(7235)	RILs	258	SSR	110	810.07	22	Shen <i>et al.,</i> 2007
9	Gh(Zhongmiansuo12)× Gh(8891)	RILs	180	SSR, AFLP, RAPD, and SRAP	132	865.20	26	Wang <i>et al.,</i> 2006
10	Gh(L-70) × Gh(L-47)	RILs	76	EST-SSR	-	-	-	Abdurakhmonov et al., 2007
11	Gh(7235) × Gh(TM-1)	RILs	207	SSR	156	1024.4	31	Shen <i>et al.,</i> 2007
12	Gh(Yumian 1) × Gh(T586)	RILs	270	SSR	19	96.2	1	Wan <i>et al.,</i> 2007
13	Gh(Deltapine) × Gh(Texas 701)	F2	251	SSR	73	650.8	17	Guo <i>et al.,</i> 2008
14	Gh× Gh	4WC	273	SSR, ESTSSR	286	2113.3	56	Qin <i>et al.,</i> 2008
15	Gh(DH962) × Gh(Jimian5)	F2	137	SRAP, SSR, RAPD and RGAP	471	3070.2	51	Lin <i>et al.,</i> 2009
16	Gh (HS 46) × Gh (MARCABUCAG8US- 1-88)	RILs	188	SSR	125	965	26	Wu <i>et al.,</i> 2009
17	Gh(Yumian 1) × Gh(T586)	RILs	270	SSR and SRAP	604	3140.9	60	Zhang <i>et al.,</i> 2009
18	Gb(Hai7124 × Gb (3- 79)	F2	124	SSR, ESTSSR, SNP	412	2108.34	52	Wang <i>et al.,</i> 2013
19	Gh (Yumian 1 × 7235)	RILs	180	SSR	1,540	2,842.06	26	Tang <i>et al.,</i> 2015
20	Gh (Yesil × Nazilli 84)	F2	94	AFLP	240	2068.5	27	Cuming <i>et al.,</i> 2015

Table 3. List of QTLs recognized in upland cotton.

S. No	Traits	Descriptor	Populatio n	Marker	(number and Type)	QTLs No.	Reference
			Туре	Siz e	216 RFLP, 139 RAPDs		Kohel <i>et al.,</i> 2001
1	Fiber quality	FS, FL, FF	F2	171	217 SSRs, 800 RAPDs UBC and 1040 OPERON	13	Zhang <i>et al.,</i> 2003
		FS	F2	186	144 AFLPs, RFLPs and 150	2	Mei <i>et al.,</i> 2004

International Journal of Nanotechnology and Allied Sciences | https://journals.psmpublishers.org/index.php/ijnas



2018; 2(2): 39-60

					SSRs		
		LY, LP, SW, NS, UQ, SF, FL, FE, FT, FF and IF	F2	120	448 RFLP	28	Zhang <i>et al.,</i> 2011
		FS,FE, FF, FU and FL	F2	200	290 SSRs and 9 AFLPs	28	Zhang <i>et al.,</i> 2005
		FS, FE, FL, FU, LP and FF	F2	117	262 RFLPs	16	Draye <i>et al.,</i> 2005
		FF	BC3F2	3,662	262 RFLPs	41	Chee <i>et al.,</i> 2005
		FL, FLU and SFC	BC3F2	3,662	95 SSRs, 72 CSR	45	Park <i>et al.,</i> 2005
		FS, FL, FF, FE	RILs	-	1378 SSRs	13	Shen <i>et al.,</i> 2005
		FL, FS, FF and FE	F2	-	4106 SSRs, AFLPs, RAPDs and SRAPs	39	Wang <i>et al.,</i> 2006
		FS, FL, FF, FMT, FE and SFI	RIL's	180	7508 SSRs, 384 SRAPs and 740 IT-ISJs	48	Zhang <i>et al.,</i> 2009
		FS, FE, FU, FL and FF	RIL's	270	16052 SSRs	13	Zhang <i>et al.,</i> 2012
		FE, FL, FS, FF and FU	CP	172	25,313 SSRs	63	Tang <i>et al.,</i> 2015
		FE, FL, FS, FF and FU	RIL's	180	123 AFLPs	62	Cuming <i>et al.,</i> 2015
		FL, FS, FE, FU and FC	F2	94	141 SSRs	43	Wu <i>et al.,</i> 2009
2	Fiber and agronomi cal	SCY, LY, LP, BW, SI, FMT, PER, WF,WT, FF, FL, FE and FS	RIL's	188	50 EST, 18 EST-SSR, 36 SSRs and 64 AFLP	36	Samer <i>et al.,</i> 2015
		BW, LP, FF.ES, FU, DFF and DFN	F2	60	834 SSRs, 437 SRAPs, 107 RAPDs, 16 REMAPs	81	He <i>et al.,</i> 2008
3	Yield and fiber	SCY, LI, SI, LY, no. of seeds per boll, FS, FL and FF	F2	69	2131 SSRs	57	Shen <i>et al.,</i> 2007
		FS, FL, FF, FE, LP, SI, NB, SCY and LY	RIL's	258	834 SSRs, 437 SRAPs, 107 RAPDs and 16 REMAPs	53	He <i>et al.,</i> 2007
		LI, SI, LY, SCY, NSB and FS	F2	69	6123 SSRs and EST-SSRs	31	Qin <i>et al.,</i> 2008
		NB, BW, SI, LP, LI, SCY, LY, FL, FS, FF, FE and FU	4WCandin bred lines	280	2675 EST-SSRs	111	Liu <i>et al.,</i> 2012
		SCY, LY, NB, BW, LP, SI, LI and FBN	RIL's and IF2	180	121 SSRs	180	Zhang <i>et al.,</i> 2013
		PH, FBN, BW, LP, LI, SI, LY, FL, FS, FE, FF and FU	G.hirsutum accessions	81	2,041 SSRs	67	Yu <i>et al.,</i> 2013

BW; Boll weight, NB: number of bolls per plant, SI: seed index, LI; Lint index, LP: lint percent, SCY; Seed cotton yield per plant, SI: seed index, LY: lint yield per plant, FS; Fiber strength, FL: fiber length, FU; Fiber Uniformity, FE: fiber elongation, FY: fiber yellowness, FF: fiber fineness, PH; Plant height, FMT: fiber maturity, FBL: fruit branch length, FBN: fruit branch number, FBA: fruit branch angle, FLU: fiber length uniformity, SFC: short fiber content, FR: fiber reflectance, SW: seed weight, NS: number of seeds per bolls, UQ: upper quartile length, SF: short fiber content, FT: fiber tenacity, IF: immature fiber content, SFI: short fiber index, NSB: number of seeds per boll, Date of 1st Flowering (DFF), Node of 1st Fruiting Branch (FFN).

CONCLUSION

Molecular Marker-Assisted technology for upland cotton Improvement involve in most specific permeative application along SSR and AFLP markers. Being costly effective, simple to manage and devoid of any radioisotope demand, SNP and simple sequence repeats molecular markers are known as the most important suitable and dependable system for DNA fingerprinting. Marker assisted selected had been successful for introgression and pyramiding major-effect genes, moreover other many dispute remain to be solved before marker assisted selection can routinely sum value for breeding very complex factors. MAS for qualitative characters aspect

most successful after deoxyribonucleic acid fingerprinting whereas for quantitative characters, insect resistance genes and genes controlling quantitative trait loci for abiotic stress tolerance, that is anticipated that application of molecular markers will remain restricted in these zones till the allele-specific markers are available and the price of molecular marker analysis is decreased. Although there had been numerous quantitative trait loci mapping studies for a broad range in upland cotton crop, comparatively a little molecular markers have actually been enforced in cotton breeding programs for cotton improvement. The scope, rate, and scale, of uptake of marker assisted selection in crop breeding program have continuously slowdown behind expectations. There are many expert and logistical components that have block the speed and compass of marker assisted selection uptake. Steady advancement and advancement in DNA markers will make it more attractive for molecular crop breeding and plant genetics and finally used in upland cotton for improvement.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interests regarding the publication of this review paper.

ACKNOWLEDGEMENTS

We are very thankful to Professor Dr. Fan Shuli and my teacher Qifeng Ma for her/ fruitful discussion on this research paper. This research project was financially sponsored by the Henan Province key R & D and extension Project (182102110048) and Key Scientific Research projects in Henan Colleges and Universities (17B180001).

AUTHOR CONTRIBUTIONS

QFM conceived to wrote the review paper, LW, XW, ZHJ investigations and interpretations of this review paper, AHJ wrote this review paper, and FS finalized the review paper, all authors read and approved the final Review paper for publication.

ABBREVIATIONS

AP-PCR= Arbitrarily primed-PCR, RAPD= Random amplified polymorphic DNA, AFLP= Amplified fragment length polymorphism, RFLP= Restriction fragment length polymorphism, CAPS= Cleaved amplified polymorphic sequence, DAF= DNA amplification fingerprinting, SNP=Single nucleotide polymorphism, SSR= Simple sequence repeats, SSCP= Single Strand Confirmation Polymorphism, SCAR= Sequence characterized amplified region, TRAP= Target region amplification polymorphism, SRAP= Sequence Related Amplified Polymorphism, RAMP= Randomly amplified micro satellite polymorphisms, REMAP= Retransposon Microsatellite Amplified Polymorphism, IRAP= Inter-retrotransposon amplified polymorphism, TD= Transposable display, MITES= Miniature Inverted Repeat transposable elements, S-SAP= Sequence-Specific amplification polymorphism, IMP= Inter-MITE polymorphism, EST= Expressed Sequence Tag, ROS= Reactive oxygen species, GWAS= Genome Wide Association Studies, RIL= Recombinant inbred line, WGS= Whole genome sequence.

REFERENCES

- Abbas, A., Iqbal, M.A., Rahman, M., Paterson, A.H., 2015. Estimating genetic diversity among selected cotton genotypes and the identification of DNA markers associated with resistance to cotton leaf curl disease. Turk. J. Bot. 3(9): 1033–1041. 10.3906/bot-1505-22.
- Abdalla, A.M., Reddy, O.U.K., El-Zik, K.M., and Pepper, A.E., 2001. Genetic diversity and relationships of diploid and tetraploid cottons revealed using AFLP. Theor. Appl. Genet., 102: 222-229.
- Agarwal, N., Shrivastava, and H., Padh, 2008. "Advances in molecular marker techniques and their applications in plant sciences," Plant Cell Reports, vol. 27, no. 4, pp. 617–631.
- Akihiro, T., Umezawa, T., Ueki, C., Lobna, B., Mizuno, K., Ohta, M., Fujimura, T., 2006. Genome wide cDNA-AFLP analysis of genes rapidly induced by combined sucrose and ABA treatment in rice cultured cells. FEBS Lett 580(25):5947–5995.
- Arunita, R., Rakshit, S., Santhy, V., Gotmare, V. P., Mohan, P., Singh, V.V., Singh, S., Singh, J., Balyan, H.S., Gupta P. K. and Bhat, S. R., 2010. Evaluation of SSR markers for the assessment of genetic diversity and fingerprinting of *Gossypium. hirsutum* accessions. J. Plant Biochem. Biotechnol. 1(9): 153-160.
- Allen, A.M., Barker, G.L.A., Wilkinson, P., Burridge, A., Winfield, M., Coghill, J., Uauy, C., Griffiths, S., Jack, P., Berry, S., Werner, P., Melichar, J.P.E., McDougall, J., Gwilliam, R., Robinson, P. and Edwards, K.J. 2013. Discovery and development of exome-based, codominant single nucleotide polymorphism markers in hexaploid wheat (*Triticum aestivum L.*). *Plant. Biotechnol. J* 1(1), 279-295.
- Altaf, J. M. C. D., Stewart, M. K., Wajahatullah, J., Zhang, and R. G., Cantrell, 1997. "Molecular and morphological genetics of a trispecies F2 population of cotton," in Proceedings of the Beltwide Cotton Conferences, vol. 1, pp. 448–452, New Orleans, La, USA.

- Atwell, S., Huang, Y.S., Vilhjalmsson, B.J., Willems, G., Horton, M., Li, Y., Meng, D., 2010. Genome-wide association study of 107 phenotypes in Arabidopsis thaliana inbred lines. Nature, 46(5), 627–631.
- Ashrafi, H., Hulse-Kemp, A.M., Wang, F., Yang, S.S., Guan, X., Jones, D.C., Matvienko, M., Mockaitis, K., Chen, Z.J. and Stelly, D.M. 2015. A Long-Read Transcriptome Assembly of Cotton and Intraspecific Single Nucleotide Polymorphism Discovery. Plant Genome. 8, 1-14.
- Bachlava, E., Taylor, C.A., Tang, S., Bowers, J.E., Mandel, J.R., Burke, J.M., and Knapp, S.J. 2012. SNP discovery and development of a high-density genotyping array for sunflower. PLoS One 7:e29814.
- Badigannavar, A., and Myers, G., 2010. Genetic analysis of AFLP markers associated with seed quality traits in upland cotton (*Gossypiumhirsutum*). In Beltwide Cotton Conferences, New Orleans, La, USA.
- Binyamin, R., M., Khan, N., Khan, and A., Khan. 2015. Application of SCAR markers linked with mung bean yellow mosaic virus disease-resistance gene in Pakistan mung bean germplasm. Genet. Mol. Res. 14: 2825-2830.
- Blackmore, T., Thomas, I., McMahon, R., Powell, W., and Hegarty, M., 2015. Genetic–geographic correlation revealed across a broad European ecotypic sample of perennial ryegrass (*Lolium perenne*) using array-based SNP genotyping. Theor. Appl. Genet., 12(8), 1917-1932.
- Blenda, A., Scheffler, J., Scheffler, B., Palmer, M., Lacape, J., Yu, J. Z., Jesudurai, C., Jung, S., Muthukumar, S., Yellambalase, P., Ficklin, S., Staton, M., Eshelman, R., Ulloa, M., Saha, S., Burr, B., Liu, S., Zhang, T., Fang, D., and Main, D., 2006. CMD: a cotton microsatellite database resource for Gossypiumgenomics. BMC Genomics, 7: 132.
- Botstein, D., White, RL., Skolnick, M., Davis, RW., 1980. Construction of a genetic linkage map in man using restriction fragment length polymorphisms. Am. J. Hum Genet., 32:314–333.
- Caetano-Anolle´s, G., Bassam, BJDNA, 1993. Amplification finger- printing using arbitrary oligonucleotide primers. App Biochem. Biotechnol. 42:189–200.
- Cavanagh, C., Morell, M., Mackay, I., and Powell, W., 2008. From mutations to MAGIC: resources for gene discovery, validation and delivery in crop plants. Curr. Opin. Plant Biol., 11, 215–221.
- Chalmers, K. J., Waugh, R., Sprent, J. I., Simons, A. J., and Powell, W., 1992. Detection of genetic variation between and within populations of Gliricidiasepium and G. maculata using RAPD markers. Heredity, 6(9): 465-472.
- Chang, Y., Guo, W.Z., Li, G.Y., Gao, F., Lin, S.S., and Zhang, T.Z., 2008. QTLs mapping for Verticillium wilt

resistance at seedling and maturity stages in *Gossypium barbadense L.* Plant Sci. 174, 290-298.

- Chagne, D., Crowhurst, R.N., Troggio, M., Davey, M.W., Gilmore, B., Lawley, C., Vanderzande, S., Hellens, R.P., Kumar, S., Cestaro, A., *et al.* 2012. Genomewide SNP detection, validation, and development of an 8K SNP array for apple. PLoS One 7:e31745.
- Chaudhary, L., Sindhu, A., Kumar, M., Kumar, R., and Saini, M., 2010. Estimation of genetic divergence among some cotton varieties by RAPD analysis. J. Plant Breed. Crop Sci., 2: 39–43.
- Chee, P. W., Draye, X., and Jiang, C. X., 2005. Molecular dissection of phenotypic variation between Gossypiumhirsutum and Gossypium barbadense (cotton) by a backcross-self approach: III. Fiber length. Theory. Appl. Genet., 111(4): 772-781.
- Chen, G., and Du, X.M., 2006. Genetic diversity of source germplasm of upland cotton in China as determined by SSR marker analysis. Acta, Genet. Sin. 33, 733–745.
- Chen, X., Wang, J., Źhu, M., Jia, H., Liu, D., Hao, L., and Guo, X., 2015. A cotton Raf-like MAP3K gene, GhMAP3K40, mediates reduced tolerance to biotic and abiotic stress in Nicotiana benthamiana by negatively regulating growth and development. Plant, Sci. 240, 10-24.
- Clarke, W.E., Higgins, E.E., Plieske, J., Wieseke, R., Sidebottom, C., Khedikar, Y., Batley, J., Edwards, D., Meng, J., Li, R., *et al.* 2016. A high-density SNP genotyping array for Brassica napus and its ancestral diploid species based on optimised selection of singlelocus markers in the allotetraploid genome. Theor. Appl. Genet., 12(9):1887–1899.
- Collard, B. C. Y., Jahufer, M. Z. Z., Brouwer, J. B., and Pang, E. C. K., 2005. An introduction to markers, quantitative trait loci (QTL) mapping and markerassisted selection for crop improvement: The basic concepts. Euphytica, 142: 169–196.
- Crowell, S., Korniliev, P., Falcao, A., Ismail, A., Gregorio, G., Mezey, J., and McCouch, S., 2016. Genome-wide association and high-resolution phenotyping link Oryza sativa panicle traits to numerous trait-specific QTL clusters. Nat. Commun., 7, 10527.
- Cuming, D. S., Altan, F., Akdemir, H., Tosun, Gurel, A., and Tanyolac, B., 2015. QTL analysis of fiber color and fiber quality in naturally green colored cotton (*GossypiumhirsutumL*.). Turkish J. Field Crop. 20(1): 49-58.
- Dahab, A. A., Saeed, M., and Mohamed, B. B., 2013. Genetic diversity assessment of cotton (*Gossypium hirsutumL*.) genotypes from Pakistan using simple sequence repeat marker. Australian, J. Crop Sci. 7(2): 261–267.
- Davey, J.W., Hohenlohe, P.A., Etter, P.D., Boone, J.Q., Catchen, J.M., and Blaxter, M.L., 2011. Genome-wide

genetic marker discovery and genotyping using nextgeneration sequencing. Nat. Rev. Genet., 12, 499-510.

- Dhruv, S., Cotton products annual report, India, 2015. USDA Foreign agricultural service, GAIN. Report Number; IN5039,
- D. D. Fang, L. L., Hinze, R. G., Percy, P. Li., D., Deng, and G. Thyssen, 2013. "A microsatellite-based genomewide analysis of genetic diversity and linkage disequilibrium in Upland cotton (*Gossypium hirsutum L.*) Cultivars from major cotton-growing countries," Euphytica, vol., 191, no. (3), 391–401, pp.
- Elshire, J. C., Glaubitz, Q., Sun. *et al.*, 2011. "A robust, simple genotyping by sequencing (GBS) approach for high diversity species," PloS. ONE. vol. 6, no. (5), Article ID e19379.
- Famoso, A.N., Zhao, K., Clark, R.T., Tung, C.W., Wright, M.H., Bustamante, C., Kochian, L.V. *et al.* 2011. Genetic architecture of aluminum tolerance in rice (Oryza sativa) determined through genome-wide association analysis and QTL mapping. PloS. Genet. 7, e1002221.
- Fang, D.D., Jenkins, J.N., Deng, D.D., McCarty, J.C., Li, P., and Wu, J.X., 2014. Quantitative trait loci analysis of fiber quality traits using a random-mated recombinant inbred population in Upland cotton (*Gossypium hirsutum L.*). BMC Genom. 15, 397.
- Fritsche-Neto, R., and Borem, A., 2012. Plant breeding for biotic stress resistance. New. York: Springer, Ganal, M.W., Durstewitz, G., Polley, A., Berard, A., Buckler, E.S., Charcosset, A., Clarke, J.D., Garner, E., Hansen, M., Joets, J., *et al.* Large maize (*Zea mays L.*) SNP genotyping array: development and germplasm genotyping, and genetic mapping to compare with the B73 reference genome. PLoS One 6:e28334.
- Geng, X., Jiang, C., Yang, J., Wang, L., Wu, X., and Wei, W., 2016. Rapid identification of candidate genes for seed weight using the SLAF-seq method in *Brassica*. *napus*. PloS. One. 11, e0147580.
- Geng, Z. Z., Gong, J. Q., Huang, and Z. C., Zhang, 1995. "Identification of difference between cotton cultivars (G. hirsutum) using the RAPD method," Jiangsu Journal of Agricultural Sciences, vol. 11, no. 4, pp. 21–24.
- Guo, W., Zhang, T., Shen, X., Yu, JZ., Kohel RJ. 2003. Development of SCAR marker linked to a major QTL for high fiber strength and its usage in molecularmarker assisted selection in upland cotton. Crop. Sci., 43:2252–2256.
- Hamilton, J.P., Hansey, C.N., Whitty, B.R., Stoffel, K., Massa, A.N., Van Deynze, A., De, Jong, W.S., Douches, D.S., and Buell, C.R., 2011. Single nucleotide poly morphism discovery in elite North American potato germplasm. BMC. Genomics. 12:302.
- Hulse-Kemp, A.M., Lemm, J., Plieske, J., Ashrafi, H., Buyyarapu, R., Fang, D.D., Frelichowski, J. *et al.* 2015. Development of a 63K SNP array for cotton and high-

density mapping of intraspecific and Inter specific populations of *Gossypium spp.* G3 Genes. Genome, Genet. 5, 1187–1209.

- Huang J, Sun M., 1999. A modified AFLP with fluorescence- labelled primers and automated DNA sequence detection for efficient fingerprinting analysis in plants. Biotechnol, Techn. 14:277–278.
- He, D., H., Lin, Z. X., and Zhang, X. L., 2008. Dissection of genetic variance of fiber quality in advanced generations from an interspecific cross of *Gossypium hirsutum* and *G. barbadense*. Pl. Breed. 127(3): 286-294.
- Ince, A. G., Karaca, M., and Onus, A. N., 2010. CAPSmicrosatellites: use of CAPS method to convert non polymorphic microsatellites into useful markers. Mol. Breed., 25(3): 491- 499.
- A.Ullah, A., Iram, M. Z., Iqbal, M., Nawaz, S., M., Hasni, and S., Jamil, 2011. "Genetic diversity analysis of Bt cotton genotypes in Pakistan using simple sequence repeat markers," Genetics, and Molecular, Research, vol. 11, no. (1), 597–605, pp.
- Ingvarsson, P.K., and Street, N.R., 2011. Association genetics of complex traits in plants. New, Phytol. 18(9), 909–922.
- Iqbal, M.J., Y., Maqsood, Z.U., Abdin, A., Manzoor, M., Hassan, and A., Jamil, 2016. SSR markers associated with proline in drought tolerant wheat germplasm. Appl. Biochem. Biotechnol. 17(8): 1042-1052.
- Jamshed, M., Jia, F., Gong, J., Palanga, K.K., Shi, Y., Li, J., Shang, H., *et al.* 2016. Identification of stable quantitative trait loci (QTLs) for fiber quality traits across multiple environments in Gossypium hirsutum recombinant inbred line population. BMC Genom. 17, 197.
- Jenkins, N, J., Jack, M. C., Martin, J. W., Russel, H., Osman, A. G., Franklin, C., and Dewayne, D., 2012. SSR Markers for Marker Assisted Selection of Root Knot nematod (Meladogyne incognita) resistant plant in cotton (*Gossypium hirsutum L*). Euphytica. 18(3): 49-54.
- Kaur, S., Francki, M., and Forster, J., 2012. Identification, characterization and interpretation of single nucleotide sequence variation in allopolyploid crop species. Plant. Biotechnol, J. 10, 125-138.
- Kalia, R., K., Rai, M. K., Kalia, S., Singh, R., and Dhawan,A. K., 2011. Microsatellite markers: an overview of the recent progress in plants, Euphytica. 177(3): 309 -334.
- Kim, S.I., and Tai, T.H., 2013. Identification of SNPs in closely related Temperate Japonica rice cultivars using restriction enzyme-phased sequencing. PloS, One. 8, e60176.
- K. Wang, Z., Wang, F., Li, 2012. "The draft genome of a diploid cotton *Gossypium raimondii*," Nature. Genetics. vol. 44, no. (10), 1098–1103, pp.

- Lacape, J. M., Nguyen, T. B., and Thibivilliers, S., 2003. A combined RFLP-SSR-AFLP map of tetraploid cotton based on a *Gossypiumhirsutum* × *Gossypiumbarbadense* backcross population.
- Lan, T. H., Cook, C. G., and Paterson, A. H., 1999. Identification of a RAPD marker linked to male fertility restoration gene in cotton (*Gossypiumhirsutum L.*). J. Agri. Genom., 1: 1-5.
- Landry, B. S., Kesseli, RV., Farrara B, Michelmore, RW., 1987. A genetic map of lettuce (*Lactuca sativa L*.) with restriction fragment length polymorphism, isozyme, disease resistance and morphological markers. Genetics, 116: 331-337.
- Li, F., Fan, G., Wang, K., Sun, F., Yuan, Y., Song, G., Li, Q., 2014a. Genome sequence of the cultivated cotton *Gossypium arboreum*. Nat. Genet.,46, 567–572.
- Li, F.G., *et al.*, 2014. Genome sequence of the cultivated cotton *Gossypium arboreum*. Nat. Genet., 46, 567–572.
- Li, X., Jin, X., Wang, H., Zhang, X., and Lin, Z., 2016. Structure, evolution, and comparative genomics of tetraploid cotton based on a high-density genetic linkage map. DNA Res. 23, 283–293.
- Li, F., Fan, G., Lu, C., Xiao, G., Zou, C., Kohel, R.J., Ma, Z., et al. 2015. Genome sequence of cultivated Upland cotton (*Gossypium hirsutum* TM-1) provides insights into genome evolution. Nat. Biotechnol., 33, 524–530.
- Li, Y.H., Zhao, S.C., Ma, J.X., Li, D., Yan, L., Li, J., Qi, X.T., et al. 2013b. Molecular footprints of domestication and improvement in soybean revealed by whole genome re-sequencing. BMC. Genom. 14, 597.
- Li, Y.H., Zhao, S.C., Ma, J.X., Li, D., Yan, L., Li, J., Qi, X.T. 2013b. Molecular footprints of domestication and improvement in soybean revealed by whole genome re-sequencing. BMC Genom. 14, 597.
- Li, Z., Wang, X., Zhang, Y., Zhang, G., Wu, L., Chi, J., and Ma, Z., 2008. Assessment of genetic diversity in glandless cotton germplasm resources by using agronomic traits and molecular markers. Front. Agric. Chin., 2: 245–252.
- Lin, C. H., Yeakley, J. M., McDaniel, T. K., and Shen, R., 2009. Medium to high-throughput SNP genotyping using Vera Code microbeads, DNA and RNA Profiling in Human Blood. Humana Press, New York, 129-142.
- Liu, R., Wang, B., and Guo, W., 2012. Quantitative trait loci mapping for yield and its components by using two immortalized populations of a heterotic hybrid in *Gossypiumhirsutum L.* Molecular, Breeding., 29(2): 297–311.
- Liu, S., Fan, C., Li, J., Cai, G., Yang, Q., Wu, J., Yi, X., *et al.* 2016. A genome- wide association study reveals novel elite allelic variations in seed oil content of Brassica napus. Theor. Appl. Genet. 129, 1203–1215.

- Liu, G., Mei, H., Wang, S., Li, X., Zhu, X., and Zhang, T., 2015a. Association mapping of seed oil and protein contents in upland cotton. Euphytica, 205, 637–645.
- Liu, Y., Wang, L., Mao, S., Liu, K., Lu, Y., Wang, J., Wei, Y., *et al.* 2015b. Genome-wide association study of 29 morphological traits in Aegilops tauschii. Sci. Rep. 5, 15562.
- Long, Y.M., Chao, W.S., Ma, G.J., Xu, S.S., and Qi, L.L., 2016. An innovative SNP genotyping method adapting to multiple platforms and throughputs. Theor. Appl. Genet., 130, 597-607.
- Lu, Q., Zhang, M., Niu, X., Wang, S., Xu, Q., Feng, Y., Wang, C., *et al.* 2015. Genetic variation and association mapping for 12 agronomic traits in indica rice. BMC. Genom. 16, 1067.
- Lu, W., Chu, X., Li, Y., Wang, C., and Guo, X., 2013. Cotton GhMKK1 induces the tolerance of salt and drought stress, and mediates defence responses to pathogen infection in transgenic Nicotiana benthamiana. PloS, One. 8, e68503.
- Ma, D., Hu, Y., Yang, C., Liu, B., Fang, L., Wan, Q., Liang, W., *et al.* 2016. Genetic basis for glandular trichome formation in cotton. Nat. Commun. 7, 10456.
- Mackay, I., and Powell, W., 2007. Methods for linkage disequilibrium mapping in crops. Trends Plant Sci. 12, 57–63.
- Maughan, P. J., Seghai, M. A., Maroof, G. R., Buss. And Huestis, G. M., 1996. Amplified fragment length polymorphism in soyabean: species diversity, inheritance and near -isogenic line analysis. Theor. Appl. Genet., 93: 392-401
- Meredith, W. R., 1992. RFLP association with varietal origin and heterosis. In: Proc. Beltwide Cotton Conf. (ed. D. Herber), Nashville, TN. pp. 607.
- Montanari, S., Saeed, M., M., Kim, Y., Troggio, M., Malnoy, M., Velasco, R., Fontana, P., Won, K., Durel, C.E., *et al.* 2013. Identification of Pyrus single nucleotide polymorphisms (SNPs) and evaluation for genetic mapping in European pear and interspecific Pyrus hybrids. PloS, One. 8:e77022.
- Mullis KB, Faloona, F., 1987. Specific synthesis of DNA in vitro via polymerase chain reaction. Methods, Enzymol. 155:350–355.
- Multani, D. S., and Lyon, B. R., 1995. Genetic fingerprinting of Australian cotton cultivars with RAPD markers. Genom., 38(5): 1005- 1008.
- Murtaza, N., 2006. Cotton genetic diversity study by AFLP markers. Electron. J. Biot., 9(4): 456- 460.
- Nie, X., Huang, C., You, C., Li, W., Zhao, W., Shen, C., Zhang, B., *et al.* 2016. Genome-wide SSR-based association mapping for fiber quality in Nationwide upland cotton inbreeds cultivars in China. BMC, Genom. 17, 352.
- Paterson, A. H., Saranga, Y., Menz, M., Jiang, C. X., and Wright, R. J., 2003. QTL analysis of genotype ×

2018; 2(2): 39-60

environment interactions affecting cotton fiber quality. Theor. Appl. Genet., 106(3): 384-396.

- Paterson, "Making genetic maps," in Genome Mapping in Plants 1996. pp. 23–39, RG Landes Company, Austin, Tex, USA; Academic Press, San Diego, Calif, USA.
- Patil, G., Do, T., Vuong, T.D., Valliyodan, B., Lee, J.D., Chaudhary, J., Shannon, J., G. and Nguyen, H.T., 2016. Genomic-assisted haplotype analysis and the development of high-throughput SNP markers for salinity tolerance in soybean. Scientific, Reports. 6, 1999.
- Prasanna, B.M., Pixley, K., Warburton, M.L., and Xie, C.-X., 2010. Molecular marker-assisted breeding options for maize improvement in Asia. Molecular, Breeding. 26, 339-356.
- Preetha, S., and Raveendren T. S., 2008. Molecular marker technology in cotton. Biotechnology and Molecular Biology Review, 3 (2): 032- 045.
- Qayyum, N., Murtaza, F. M., Azhar, and W., Malik, 2009. "Biodiversity and nature of gene action for oil and protein contents in *Gossypium hirsutum L.* estimated by SSR markers," Journal of Food, Agricul, and Environment, vol. 7, no. 2, pp. 590–593.
- Rafalski, J. A., 1997. Randomly amplified polymorphic DNA (RAPD) analysis. In: DNA markers: Protocols, Applications and Overviews (eds. G.C.Anolles and P.M. Gresshoff), Wiley-Liss, Inc. USA, p. 364.
- Rahman, M., Yasmin, T., Tabassum, N., Ullah, I., Asif, M., and Zafar, Y., 2008. Studying the extent of genetic diversity among Gossypium arboretum L. genotypes/cultivars using DNA finger printing technology. Genet.Resour. Crop Evol. 55: 331-339.
- Rahman, M., Hussain, D., and Zafar, Y., 2002. Estimation of genetic divergence among elite cotton cultivarsgenotypes by DNA finger printing technology. Crop Sci., 42: 2137-2144.
- Rana, M. K., Singh, V. P., and Bhat, K. V., 2005. Assessment of genetic diversity in upland cotton (*GossypiumhirsutumL*.) breeding lines by using amplified fragment length polymorphism (AFLP) markers and morphological characteristics. Genet.Resour. Crop Evol. 52: 989–997.
- Rana, M. K., Singh, V. P., and Bhat, K. V., 2004. Assessment of genetic diversity in upland cotton (GossypiumhirsutumL.) breeding lines by using amplified fragment length polymorphism (AFLP) markers and morphological characteristics. Genet.Resour. Crop Evol. 52: 989–997.
- Rasheed, A., Wen, W., Gao, F.M., Zhai, S., Jin, H., Liu, J.D., Guo, Q., Zhang, Y.J., Dreisigacker, S., Xia, X.C., and He, Z.H., 2016. Development and validation of KASP assays for functional genes underpinning key economic traits in wheat. Theor. Appl, Genet. 129, 1843-1860.

- Rasheed, A., X., Xia, T., Mahmood, U.M., Quraishi, A., Aziz, H., Bux, Z., Mahmood, J.I., Mirza, A., Mujeeb-Kazi, and Z. He., 2016. Comparison of economically important loci in landraces and improved wheat cultivars from Pakistan. Crop, Sci. 56: 287-301.
- Rakshit, A., Rakshit, S., and Singh, J., 2010. Association of AFLP and SSR Markers with agronomic and fiber quality traits in *Gossypium hirsutum* L. J. Genet., 89(2): 155-162.
- Reinisch, J., Dong, J. M., Brubaker, C. L., Stelly, D. M., Wendel, J. F., and Paterson, A. H., 1994, Int.J.Curr.Microbiol.App.Sci 2017. 6(5): 2627-2644 2642 A detailed RFLP map of cotton, Gossypiumhirsutum × Gossypiumbarbadense: chromosome organization and evolution in a disomic polyploid genome. Genet. 138(3): 829-884.
- Remington, D.L., Thornsberry, J.M., Matsuoka, Y., Wilson, L.M., Whitt, S.R., Doebley, J., Kresovich, S., 2001. Structure of linkage disequilibrium and phenotypic associations in the maize genome. Proc. Natl Acad. Sci. USA, 98, 11479–11484.
- Richard, and J. S., Beckmann, 1995. "How neutral are synonymous codon mutations?" Nature genetics, vol. 10, no. 3, article 259.
- Rong, J., Abbey, C., Bowers, J. E., Brubaker, C. L., Chang, C., Chee, P. W., Delmonte, T. A., Ding, X., Garza, J. J., Marler, B. S., Park, C., Pierce, G., J., Rainey, K. M., Rastogi, V. K., Schulze, S. R., Trolinder, N. L., Wendel, J. F., Wilkins, T. A., Williams-Coplin, T. D., Wing, R. A., Wright, R. J., Zhao, X., Zhu, L. and Paterson, A. H., 2004. A 3347-locus genetic recombination map of sequence-tagged sites reveals features of genome organization, transmission and evolution of cotton (Gossypium). Genetics, 166: 389-417.
- Saintenac, C., Jiang, D., and Akhunov, E.D., 2011.Targeted analysis of nucleotide and copy number variation by exon capture in allotetraploid wheat genome. Genome, Biol. 12, R88.
- Saidou, A.A., Thuillet, A.C., Couderc, M., Mariac, C., and Vigouroux, Y., 2014. Association studies including genotype by environment interactions: Mprospects and limits. BMC, Genet. 15, 1–12.
- Saranga.Y., Menz, M., Jiang, C. X., Wright, R. J., Yakir, D., and Paterson, A. H., 2001. Genomic dissection of genotype x environment interactions conferring adaptation of cotton to arid conditions. Genom. Res., 11(12): 1988-1995.
- Saranga, Y., Paterson, A.H. and Levi, A. (2009) Bridging classical and molecular genetics of abiotic stress resistance in cotton. In: Genetics and Genomics of Cotton pp. 337-352. Springer.
- Semagn, K., Babu, R., Hearne, S., and Olsen, M., 2014. Single nucleotide Polymorphism genotyping using competitive Allele Specific PCR (KASP): Over view of

the technology and its application in crop improvement. Mol. Breed. 33; 1-14.

- Semagn, K., Babu, R., Hearne, S., and Olsen, M., 2014. Single nucleotide polymorphism genotyping using Kompetitive Allele Specific PCR (KASP): Overview of the technology and its application in crop improvement. Molecular, Breeding. 33, 1-14.
- Shaheen, M., Asif, and Y., Zafar, 2009. "Single nucleotide polymorphism analysis of MT-SHSP gene of Gossypium arboreum and its relationship with other diploid cotton genomes, G. hirsutum and Arabidopsis thaliana," Pakistan Jour, of Botany, vol. 41, no. 1, pp. 177–183.
- Shi, A., Kantartzi, S, M., and Chen, P., 2010. Development of ISSR PCR markers for diversity study in dogwood (Cornus spp.). Agril. And Biol. J. of North America. 1: 189-194.
- Shen, X., Guo, W., and Zhu, X., 2005. Molecular mapping of QTLs for fiber qualities in three diverse lines in Upland cotton using SSR markers, Molecular. Breeding, 15(2):169–181.
- Shen, X. L., Guo, W., Lu, Q., Zhu, X., Yuan, Y., and Zhang, T., 2007. Genetic mapping of quantitative trait loci for fiber quality and yield trait by RIL approach in upland cotton. Euphytica, 155 (3): 371-380.
- Shi, J., Zhang, L., An, H., Wu, C., and Guo, X., 2011. GhMPK16, a novel Stress-responsive group D MAPK gene from cotton is involved in disease resistance and drought sensitivity. BMC, Mol. Biol. 12, 22.
- Shu, B., K., Fenling, Z. Y., Yao, Z. G., Mei, Z. Q., Yuan, and Gang, W. X., 2001. Genetic diversity analysis of representative elite cotton varieties in three main cotton regions in China by RAPD and its relation with agronomic characteristics. Scientia AgriculturaSinica, 34: 597-603.
- Singh, N., Jayaswal, P.K., Panda, K., Mandal, P., Kumar, V., Singh, B., Mishra, S., Singh, Y., Singh, R., Rai, V., *et al.* 2015. Single-copy gene based 50 K SNP chip for genetic studies and molecular breeding in rice. Sci. Rep. 5:11600.
- Smulders, M., and G., De, Klerk, 2011. Epigenetics in plant tissue culture. Plant, Growth, Regul. 63 (2): 137-146.
- Sun, X., Liu, D., Zhang, X., Li, W., Liu, H., Hong, W., Jiang, C., Guan, N., Ma, C., Zeng, H., *et al.* 2013. SLAF-seq: an efficient method of largescale de novo SNP discovery and genotyping using high-throughput sequencing. PloS, One. 8:e58700.
- Tatineni, V., Cantrell, R. G., and Davis, D. D., 1996. Genetic diversity in elite cotton germplasm determined by morphological characteristics and RAPDs. Crop Sci., 36(1): 186-192.
- Tautz D, Renz M., 1984. Simple sequences are ubiquitous repetitive components of eukaryotic genomes. Nucleic, Acids Res. 12:4127–4138.

- Thomson, M.J., 2014. High-Throughput SNP Genotyping to Accelerate Crop Improvement. Plant, Breed, and Biotech. 2, 195-212.
- Truong, H.T., Ramos, A.M., Yalcin, F., de Ruiter, M., van der Poel, H.J., Huvenaars, K.H., Hogers, R.C., van Enckevort, L.J., Janssen, A., van Orsouw, N.J., and van Eijk, M.J., 2012. Sequence-based genotyping for marker discovery and co-dominant scoring in germplasm and populations. PloS, One. 7, e37565.
- Ulloa, and W. R. Meredith, Jr., 2000. "Genetic linkage map and QTL analysis of agronomic and fiber traits in an intraspecific population," Journal of Cotton Science, vol. 4, no. 3, pp. 161–170.
- Varshney, A., Graner, and M. E., Sorrells, 2005. "Genic microsatellite markers in plants: features and applications," Trends in Biotechnology, vol. 23, no. 1, pp. 48–55.
- Vos, P., Hogers, R., and Bleeker, M., 1995. AFLP: a new technique for DNA fingerprinting. Nucleic Acids Res., 23(21): 4407-4414.
- Vos, P.G., Uitdewilligen, J.G., Voorrips, R.E., Visser, R.G., and van Eck, H.J., 2015. Development and analysis of a 20K SNP array for potato (*Solanum tuberosum*): an insight into the breeding history. Theor. Appl. Genet. 128:238 7-2 401.
- Verde, I., Bassil, N., Scalabrin, S., Gilmore, B., Lawley, C.T., Gasic, K., Micheletti, D., Rosyara, U.R., Cattonaro, F., Vendramin, E., 2012. Development and evaluation of a 9K SNP array for peach by internationally coordinated SNP detection and validation in breeding germplasm. PloS, One. 7:e35668.
- Wajahatullah, M. K., and Stewart, J. M., 1997. Genomic affinity among Gossypium subgenus Sturtia species by RAPD analysis.In Proceeding of the Belt wide Cotton Conference, National Cotton Council, p. 452, Memphis, Tenn, USA.
- Wang, C. B., Guo, W. Z., Cai, C. P., and Zhang, T. Z., 2006. Characterization, development and exploitation of EST derived microsatellites in *Gossypium raimondii* Ulbrich. Chin. Sci. Bull., 51: 557-561.
- Wang, H., Huang, C., Guo, H., Li, X., Zhao, W., Dai, B., Yan, Z., *et al.* 2015a. QTL mapping for fiber and yield traits in upland cotton under multiple environments. PLoS ONE, 10, e0130742.
- Wang, H., Jin, X., Zhang, B., Shen, C., and Lin, Z., 2015b. Enrichment of an intraspecific genetic map of upland cotton by developing markers using parental RAD sequencing. DNA Res. 22, 147–160.
- Wang, Z., Wang, F., Li, 2012. "The draft genome of a diploid cotton Gossypium raimondii," Nature Genetics, vol. 44, no. 10, pp. 1098–1103.
- Wang, M., Yan, J., Zhao, J., Song, W., Zhang, X., Xiao, Y., and Zheng, Y., 2012b. Genome-wide association study

2018; 2(2): 39-60

(GWAS) of resistance to head smut in maize. Plant Sci. 196, 125–131.

- Wang, H., Huang, C., Guo, H., Li, X., Zhao, W., Dai, B., Yan, Z., 2015a. QTL mapping for fiber and yield traits in upland cotton under multiple environments. PLoS ONE, 10, e0130742.
- Wang, H., Jin, X., Zhang, B., Shen, C., and Lin, Z., 2015b. Enrichment of an intraspecific genetic map of upland cotton by developing markers using parental RAD sequencing. DNA Res. 22, 147–160.
- Wen, W., Li, D., Li, X., Gao, Y., Li, W., Li, H., Liu, J. 2014. Metabolomebased genome-wide association study of maize kernel leads to novel biochemical insights. Nat. Commun. 5, 3438.
- Wendel J, F., Grover, C. E., 2015. "Taxonomy and evolution of the cotton genes, *Gossypium*,"*Cotton* eds Fang, D. D., Percy, R. G., editors. (Madison, WI: American Society of Agronomy Inc.) 25–44.
- Williams, JGK., Kubelik, AR., Livak, KJ., Rafalski, JA., 1991. Tingey SV DNA polymorphisms amplified by arbitrary primers are usefll as genetic markers. Nucleic, Acids, Res. 18:6531–6535.
- Williams, J., Kubelik, A., Liviak, J. L., Rafalski, J. A., Tingey, S. V., 1990. DNA polymorphisms amplified by random primers are useful as genetic markers. Nucleic, acid, Res., 18: 6531- 6535.
- Wright, P. M., Thaxton, K. M., El-Zik, and A. H., Paterson, 1998. "D-subgenome bias of Xcm resistance genes in tetraploid Gossypium (cotton) suggests that polyploid formation has created novel avenues for evolution," Genetics, vol. 149, no. 4, pp. 1987–1996.
- Wu, J., Li, L.T., Li, M., Khan, M.A., Li, X.G., Chen, H., Yin, H., and Zhang, S.L, 2014. High-density genetic linkage map construction and identification of Fruit-related QTLs in pear using SNP and SSR markers. J. Exp. Bot. 65:5771-5781.
- Xu, C., Ren, Y., Jian, Y., Guo, Z., Zhang, Y., Xie, C., Fu, J., Wang, H., Wang, G., Xu, Y., Li, P., and Zou, C., 2017b. Development of maize 55 K SNP array with improved genome coverage for molecular breeding. Mol, Breed. 37, 20.
- Xu, X. L., Zhang, and Y. C., Nie, 2001. "Genetic diversity evaluation of cultivars (G. hirsumtum L.) from the Changjiang river valley and Tellow river valley by RAPD markers," Acta Genetica Sinica, vol. 28, no. 7, pp. 683–690.
- Xu, X., Xu, R., Zhu, B., Yu, T., Qu, W., Lu, L., Xu, Q., Qi, X., and Chen, X., 2014b. A high-density genetic map of cucumber derived from Specific Length Amplified Fragment sequencing (SLAF-seq). Front. Plant, Sci. 5, 768.
- Xu, L., Hu, K., Zhang, Z., Guan, C., Chen, S., Hua, W., Li, J., 2016. Genome-wide association study reveals the genetic architecture of flowering time in rapeseed (Brassica napus L.). DNA Res. 23, 43–52.

- Xu, Q. H., Zhang, X. L., and Nie, Y. C., 2001. Genetic diversity evaluation of cultivars (*G. hirsumtumL*.) from the Changjiang river valley and Tellow river valley by RAPD markers. Acta Genet. Sin. 28: 683–690.
- Yang, H., Tao, Y., Zheng, Z., Li, C., M.W., and Howieson, J.G., 2012. Application of next-generation sequencing for rapid marker development in molecular plant breeding: a case study on anthracnose disease resistance in Lupinus angustifolius L. BMC, Genomics. 13, 318.
- Yang, N., Lu, Y., Yang, X., Huang, J., Zhou, Y., Ali, F., Wen, W., 2014. Genome wide association studies using a new nonparametric model reveal the genetic architecture of 17 agronomic traits in an enlarged maize association panel. PLoS Genet. 10, e1004573.
- Yu, J., W., Yu, S., X., Lu, C. R., Wang, W., Fan, S. L., Song, M. Z., Lin, Z. X., Zhang, X. L., and Zhang, J. F., 2007. High-density linkage map of cultivated allotetraploid cotton based on SSR, TRAP, SRAP and AFLP markers. J. Integr Plant Biol., 49: 716-724.
- Yu, Z. H., Park, Y. H., Lazo, G. R., and Kohel, R. J., 1997. Molecular Mapping of the Cotton Genome.Agron, Abstracts, ASA, Madison, Wis, USA.
- Yu, R. J., Kohe, D. D., Fang, 2012. "A high-density simple sequence repeat and single nucleotide polymorphism genetic map of the tetraploid cotton genome," G3: Genes, Genomes, Genetics. vol. 2, no. 1, pp. 43–58.
- Yu, J. W., Yu, S. X., Lu, C. R., Wang, W., Fan, S. L., Song, M. Z., Lin, Z. X., Zhang, X. L., and Zhang, J. F., 2007. High-density linkage map of cultivated allotetraploid cotton based on SSR, TRAP, SRAP and AFLP markers. J. Integr Plant Biol., 49: 716-724.
- Y. Zhang, X. F., Wang, Z. K., Li, G. Y., Zhang, and Z. Y., Ma, 2011. "Assessing Genetic Diversity of cotton cultivars using genomic and newly developed expressed sequence tag-derived microsatellite markers," Genetics, and Molecular, Research. vol. 10, no. (3), 1462–1470, pp.
- Zhang, Y., Wang, X, F., Li, Z. K., Zhang, G. Y., and Ma, Z. Y., 2011. Assessing genetic diversity of cotton cultivars using genomic and newly developed expressed sequence tag-derived microsatellite markers, Genetics, and Molecular Res., 10(3): 1462–1470.
- Zhang, T., Hu, Y., Jiang, W., Fang, L., Guan, X., Chen, J., Zhang, J., *et al.* 2015. Sequencing of allotetraploid cotton (*Gossypium hirsutum L.* acc. TM-1) provides a resource for fiber improvement. Nat. Biotechnol. 33,531-537.
- Zhang, Z., Shang, H., Shi, Y., Huang, L., Li, J., Ge, Q., Gong, J., *et al.* 2016. Construction of a high-density genetic map by specific locus amplified fragment sequencing (SLAF-seq) and its application to Quantitative Trait Loci (QTL) analysis for boll weight in upland cotton (*Gossypium hirsutum*). BMC, Plant Boil. 16, 79.

- Zhang, T., Qian, N., and Zhu, X., 2013. Variations and transmission of QTL alleles for yield and fiber qualities in upland cotton cultivars developed in China, PLoS ONE, 8(2): ID e57220.
- Zhang, Y., Yuan, J., Yu, W., Guo, and R. J., Kohel, 2003. "Molecular tagging of a major QTL for fiber strength in upland cotton and its marker-assisted selection," Theoretical and Applied Genetics, vol. 106, no. 2, pp. 262–268.
- Zhang, M., McCarty, J., CJenkins, J. N., and Saha, S., 2002. Assessment of day-neutral backcross populations of cotton using AFLP markers. J. Cotton Sci., 6(2): 97-103.
- Zhang, Y., Yuan, J., Yu, W., Guo, and R. J., Kohel, 2003. "Molecular tagging of a major QTL for fiber strength in upland cotton and its marker-assisted selection," Theoretical and Applied Genetics, vol. 106, no. 2, pp. 262–268.
- Zhang, Y. Lu, R. G., Cantrell, and E., Hughs, 2005. "Molecular marker diversity and field performance in commercial cotton cultivars evaluated in the southwestern USA," Crop Science, vol. 45, no. 4, pp. 1483–1490.
- Zhang, Z. S., Xiao, Y. H., and Luo, M., 2005. Construction of a genetic linkage map and QTL analysis of fiberrelated traits in upland cotton (Gossypiumhirsutum L.). Euphytica, 144(1-2): 91-99.
- Zhang, K., Zhang, J., and Ma, J., 2012. Genetic mapping and quantitative trait locus analysis of fiber quality traits using a three-parent composite population in upland cotton (*Gossypiumhirsutum L*.). Molec, Breed. 29(2): 335–348.
- Zhang, Y., Yuan, J., Yu, W., Guo, and R. J., Kohel, 2003. "Molecular tagging of a major QTL for fiber strength in upland cotton and its marker-assisted selection," Theor, and Applied Genetics, vol. 106, no. 2, pp. 262–268.
- Zhao, S., Fung-Leung, W.P., Bittner, A., Ngo, K., and Liu, X., 2014. Comparison of RNA-Seq and Microarray in Transcriptome Profiling of Activated T Cells. PLoS One 9, e78644.
- Zhao, K., Tung, C.W., Eizenga, G.C., Wright, M.H., Ali, M.L., Price, A.H., Norton, G.J., 2011. Genome-wide association mapping reveals a rich genetic architecture of complex traits in Oryza sativa. Nat. Commun. 2, 467.
- Zhao, K., Aranzana, M.J., Kim, S., Lister, C., Shindo, C., Tang, C., Toomajian, C., 2005. An Arabidopsis example of association mapping in structured samples. PLoS Genet. 3, e4.
- Zhong, J. C., McCarty, J. N., Jenkins, and S., Saha, 2002. "Assessment of day-neutral backcross populations of cotton using AFLP markers, Jour. of Cotton Science, vol. 6, no. 2, pp. 97–103.

Zhu, Q.H., Zhang, J., Liu, D., Stiller, W., Liu, D., Zhang, Z., Llewellyn, D., 2016. Integrated mapping and characterization of the gene underlying the okra leaf trait in Gossypium hirsutum L. J. Exp. Bot. 67, 763– 774.

2018; 2(2): 39-60